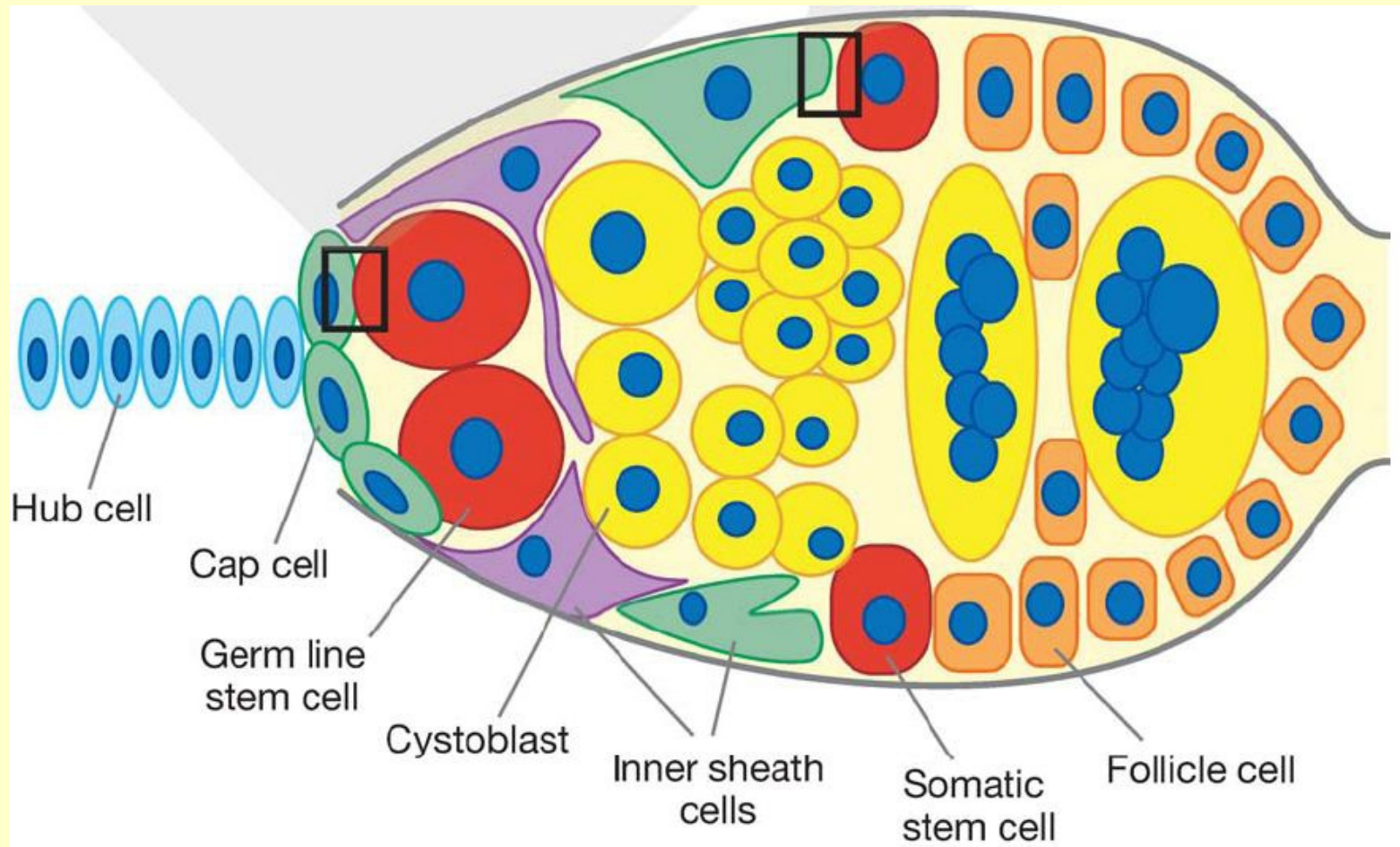


Genomics, Bioinformatics & Medicine

<http://biochem158.stanford.edu/>

Stem Cells

<http://biochem158.stanford.edu/Stem%20Cells.html>



Doug Brutlag

Professor Emeritus of Biochemistry & Medicine

Stanford University School of Medicine

Causal Mutation Homework Assignment

Most of the SNP variations associated with diseases in genome-wide association studies do not cause the disease, but instead, these SNPs serve as genetic markers that are linked to genes which are involved in the disease. Ongoing research is attempting to sequence these genes in patients and in controls to find the actual variations in these genes that do in fact, cause the disease.

For this assignment I would like you to choose a simple Mendelian inherited disease other than those mentioned in class (Huntingtons, diabetes, Parkinsons, cystic fibrosis, sickle cell, etc.) and describe what is known about the genetic variations that cause that disease.


You may search [OMIM](#), [dbSNP](#), [dbVAR](#), [HGMD](#), [HGVS](#), [ClinVar](#), [SwissVar](#) and other database of genome variations that are associated with specific diseases to find an example of the kinds of mutations associated with the disease. Please describe how each of these variations cause the disease.

Is it by:

- 1) mutating the coding region of the protein
- 2) altering the gene expression by affecting the promoter
- 3) altering gene expression by affecting a transcription factor binding site
- 4) altering gene expression indirectly by mutating a transcription factor itself
- 5) altering copy number, hence changing gene expression levels
- 6) altering other regulatory sites (miRNA targets)
- 7) altering splice signals

etc.

Often there will be several types of mutations that can cause the disease. Please comment on all types that are known for your chosen disease.



HumBio 157

The Biology of Stem Cells

HUMBIO 157: The Biology of Stem Cells (DBIO 257)

The role of stem cells in human development and potential for treating disease. Guest lectures by biologists, ethicists, and legal scholars. Prerequisites: HumBio 2A and 3A, or the equivalent in the BioCore in Biological Sciences.

Terms: Spr | Units: 3 | UG Reqs: WAY-SMA | Grading: Letter or Credit/No Credit

Instructors: Fuller, M. (PI) ; Nusse, R. (PI)

[Schedule for HUMBIO 157](#)

2014-2015 Spring

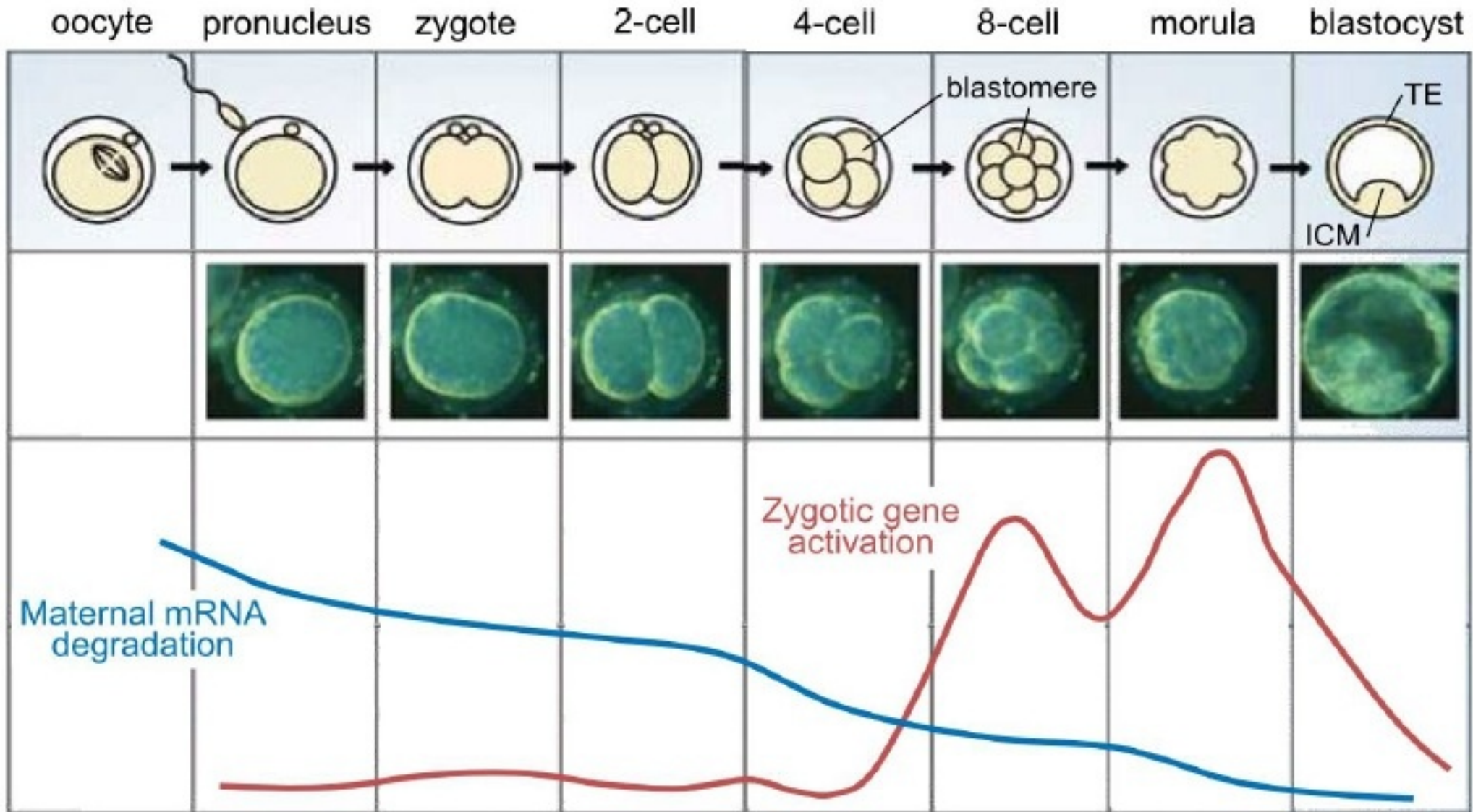
HUMBIO 157 | 3 units | UG Reqs: WAY-SMA | Class # 20511 | Section 01 | Grading: Letter or Credit/No Credit | LEC |

Students enrolled: 28

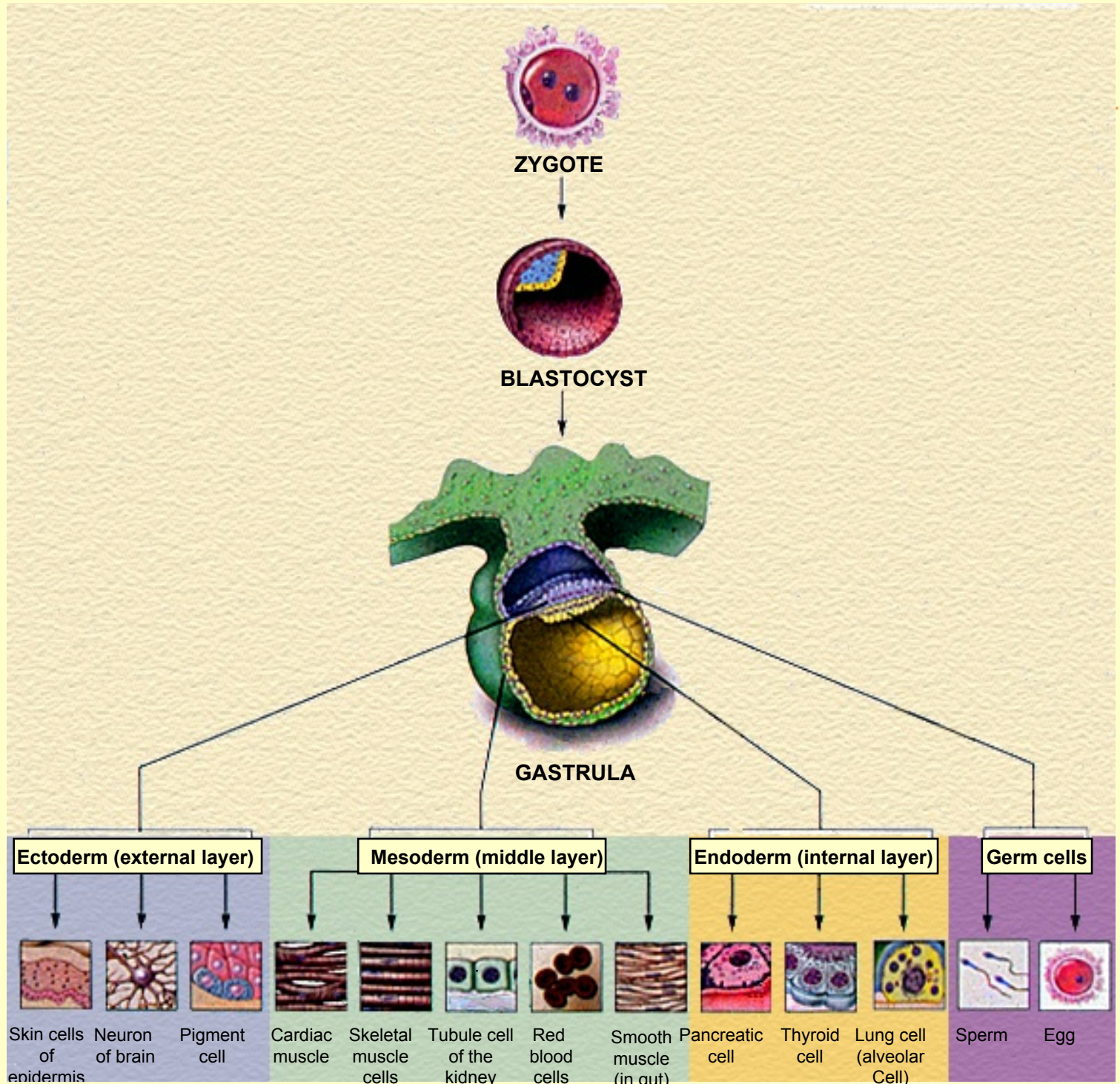
03/30/2015 - 06/03/2015 Tue, Thu 2:15 PM - 3:45 PM at [Econ 140](#) with Fuller, M. (PI); Nusse, R. (PI)

Instructors: Fuller, M. (PI); Nusse, R. (PI)

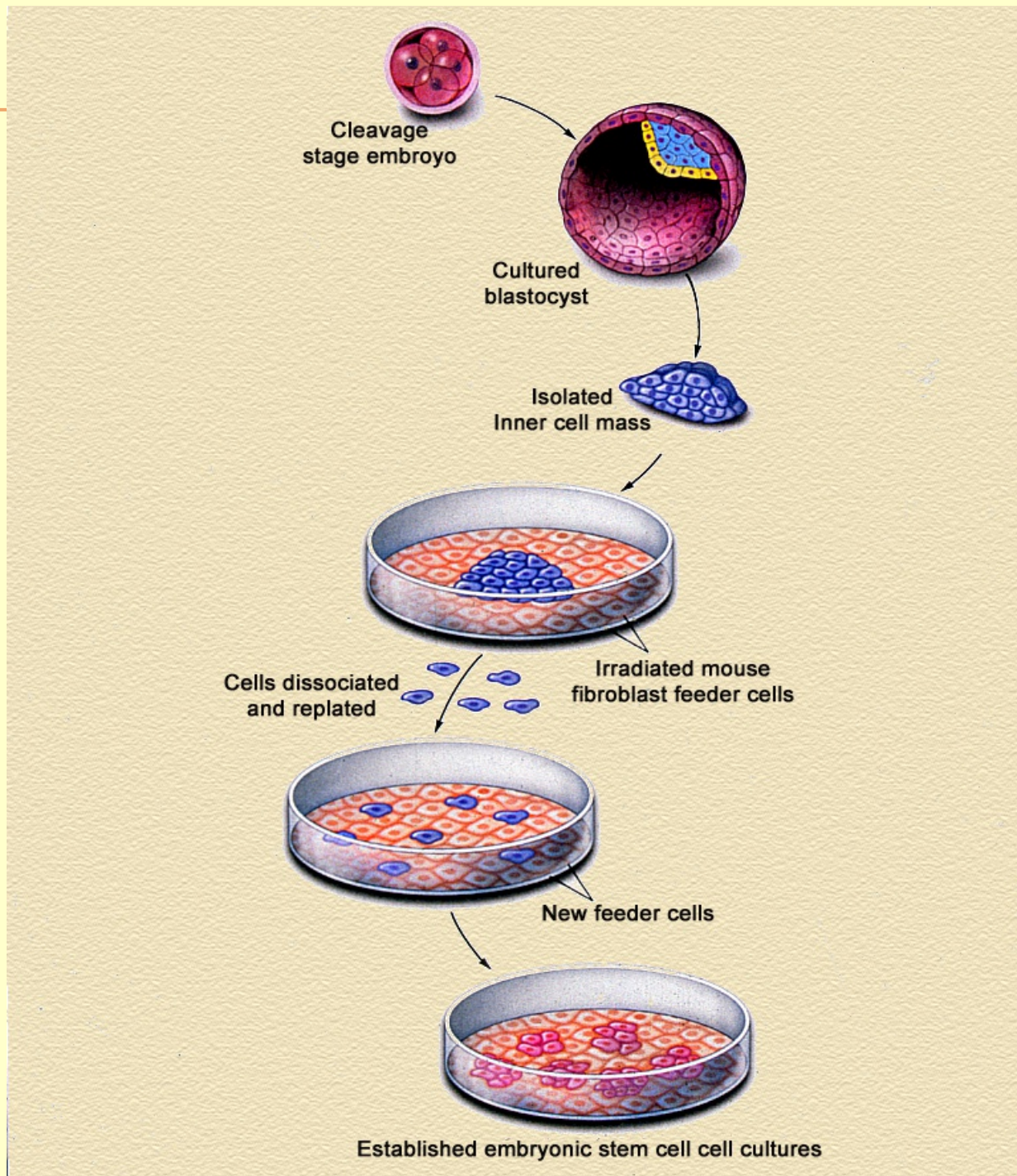
Early Embryo Development



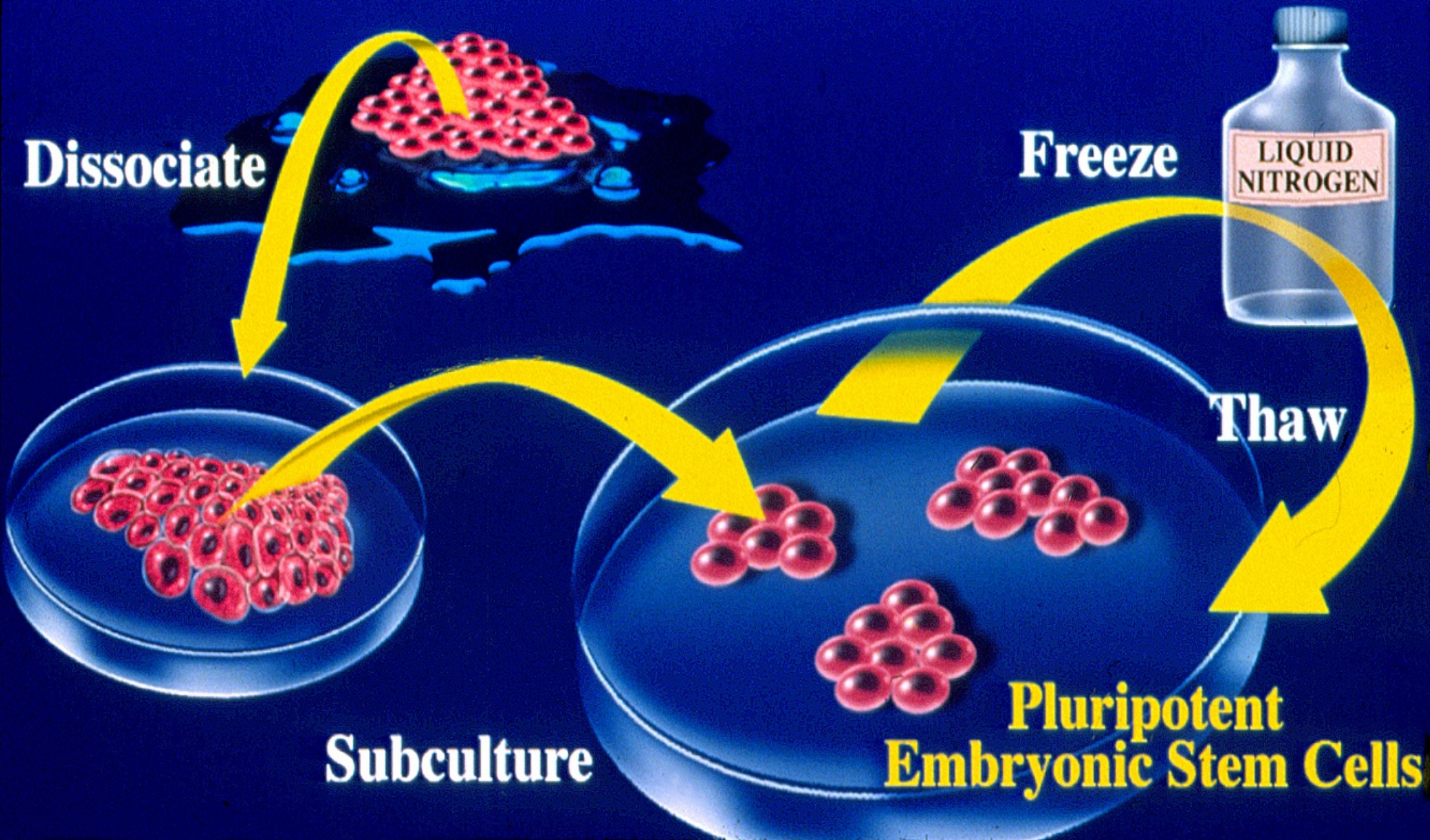
Differentiation of Human Tissues



Embryonic Stem Cell Cultures

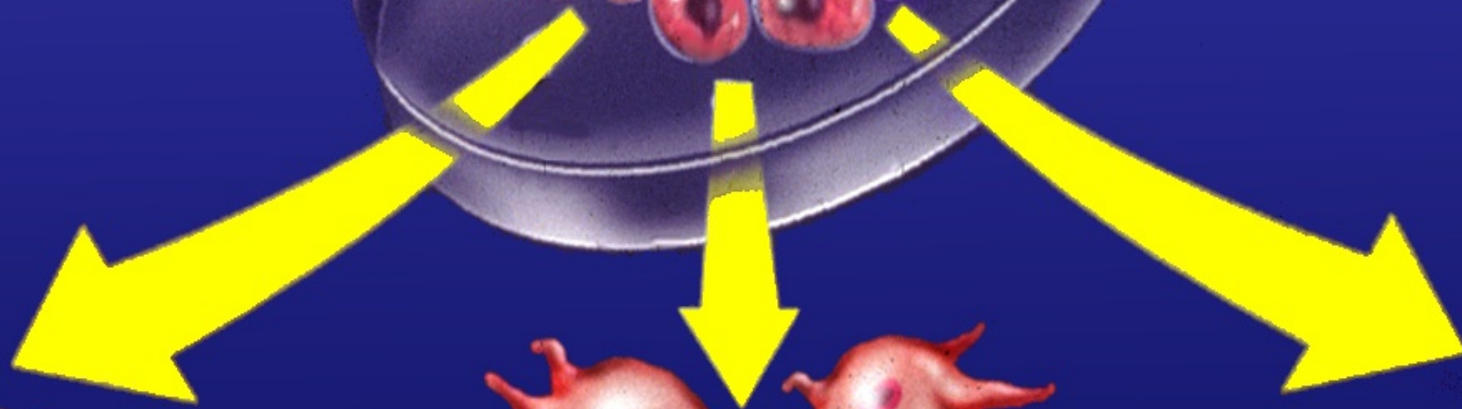
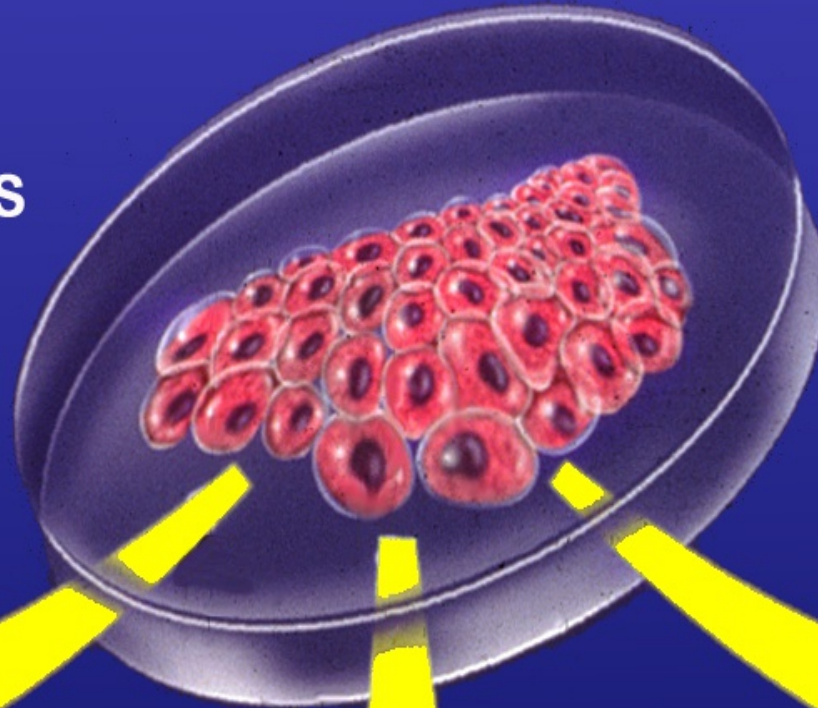


Inner Cell Mass Cells Continue to Proliferate Indefinitely in Culture

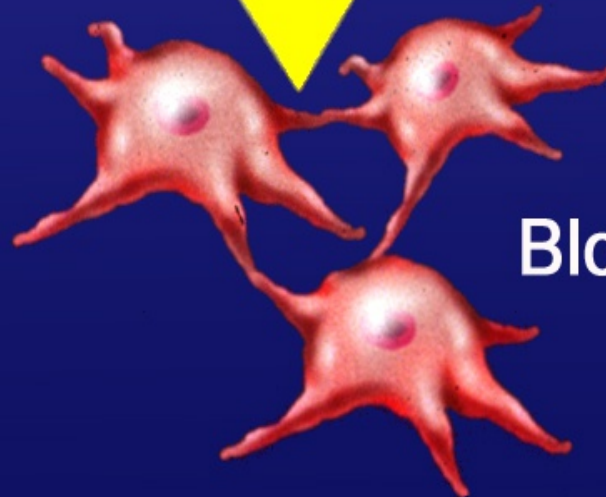


Pluripotent Stem Cells Differentiate into many Cell Types

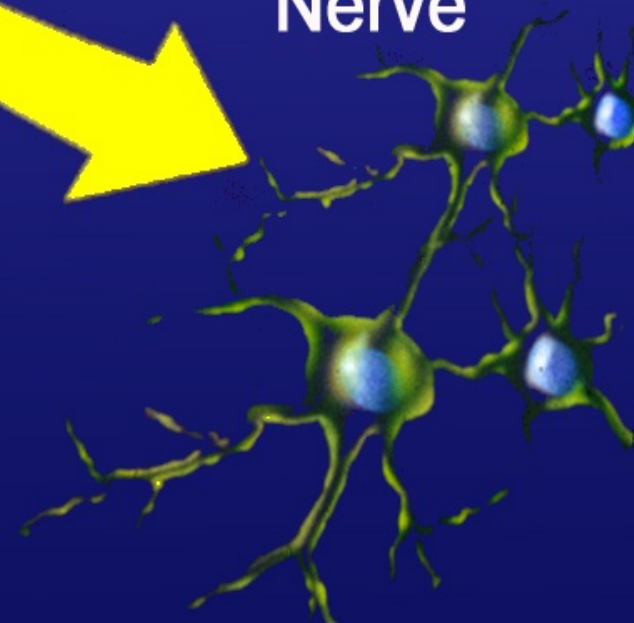
Add different growth factors



Muscle



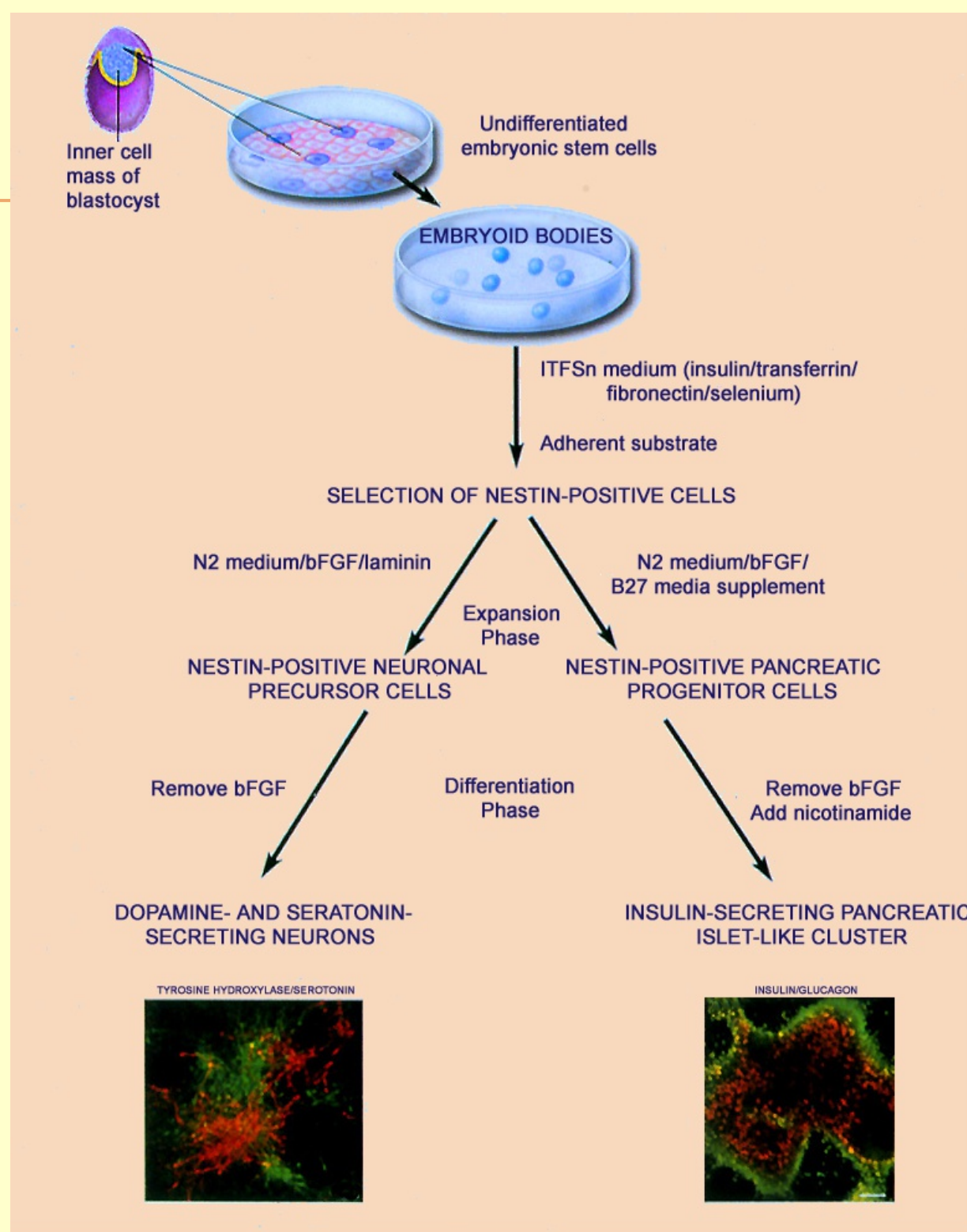
Blood



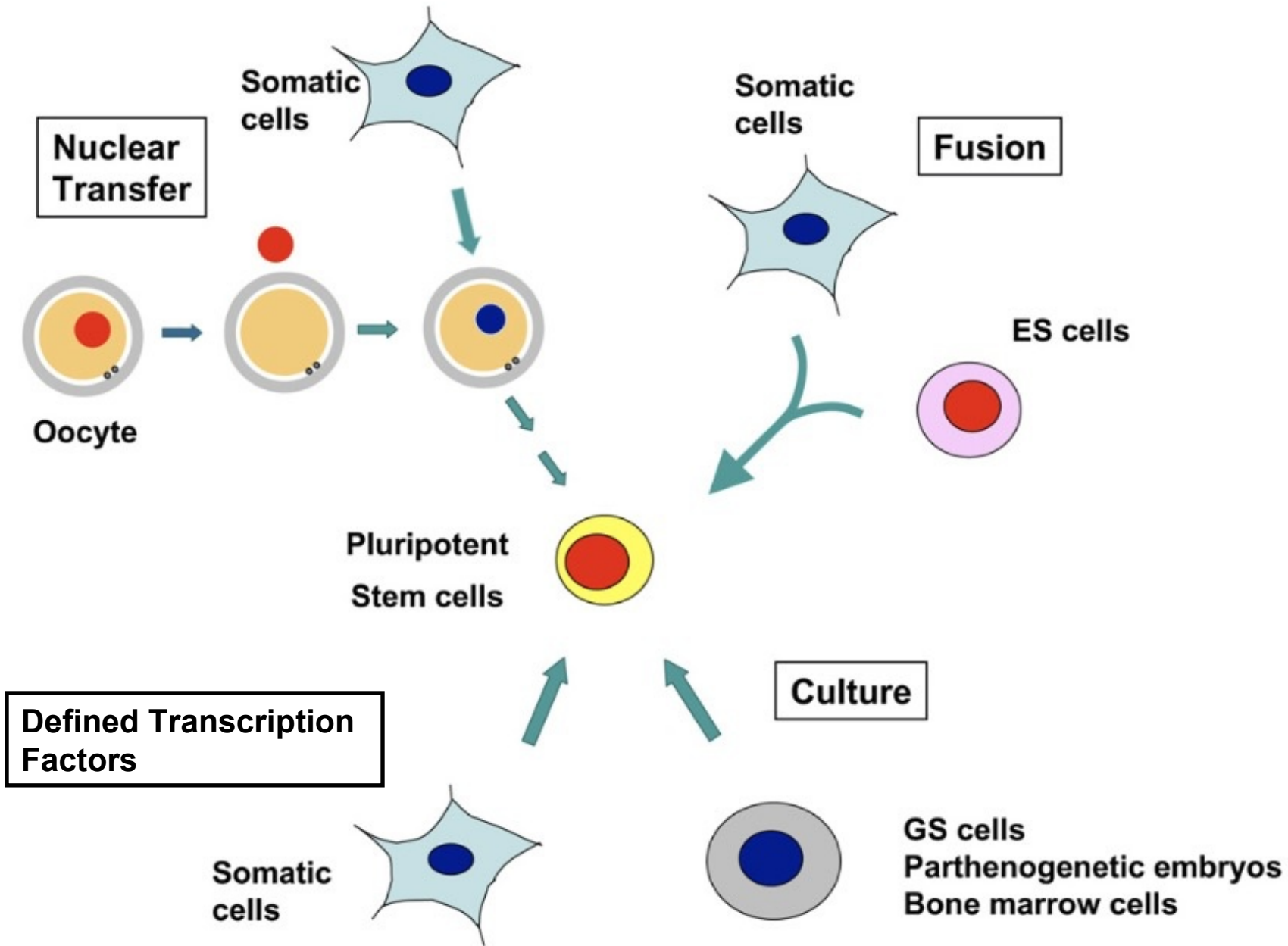
Nerve

Basic Problems of Stem Cell Therapy

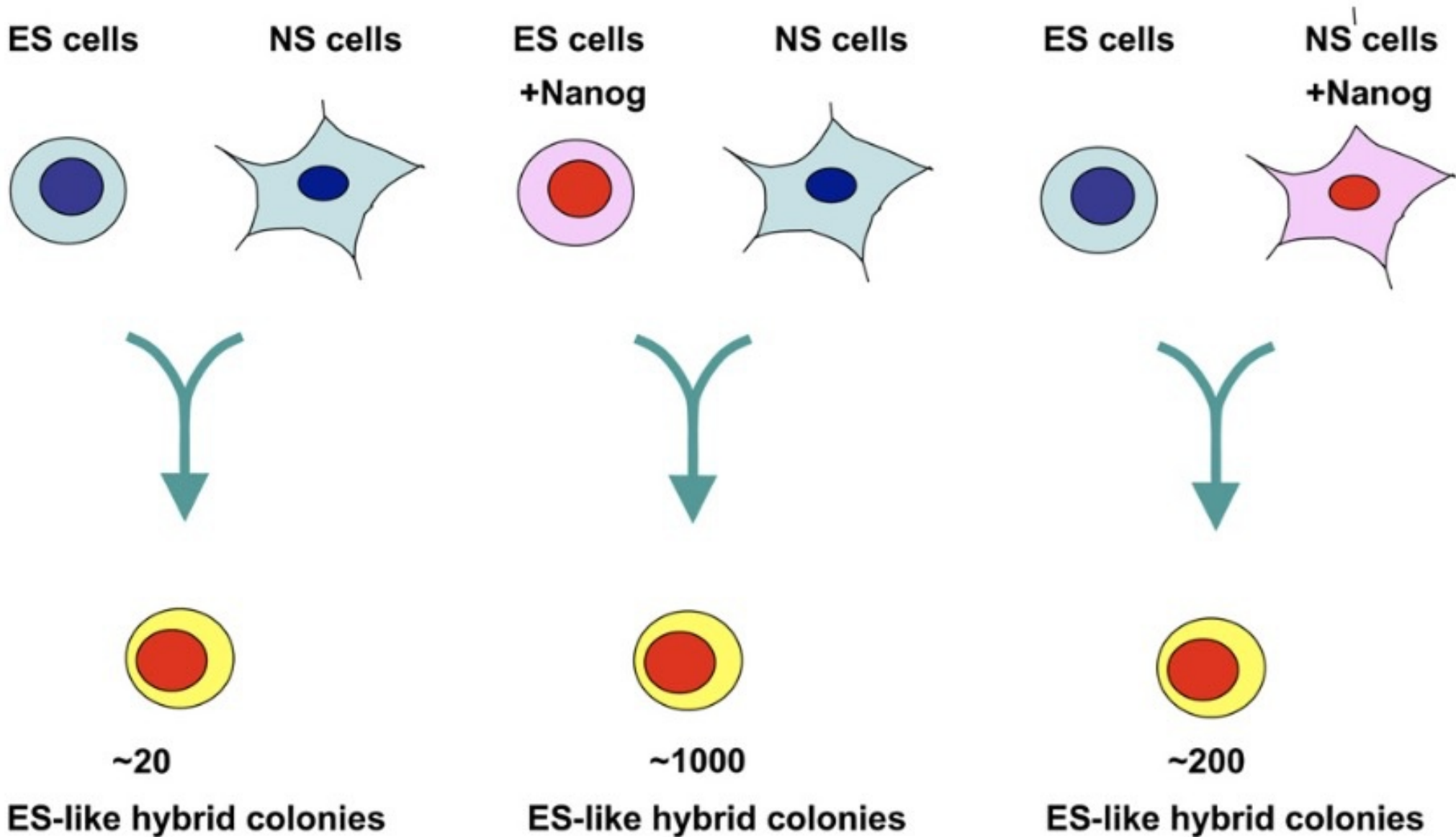
- **HOW TO DIRECT DIFFERENTIATION OF CELLS DOWN SPECIFIC PATHWAYS?**
e.g. all into muscle or all into nerve; different “cocktails” of growth factors
- **HOW TO OVERCOME IMMUNE REJECTION?**
e.g. alter histocompatibility genes; therapeutic cloning for “customized” lines
- **HOW TO MAKE AN ORGAN?**
e.g. combine different cell types in three dimensional arrangements.



Methods to Generate Pluripotent Stem Cells



Nanog-Mediated Enhancement of Reprogramming by Fusion

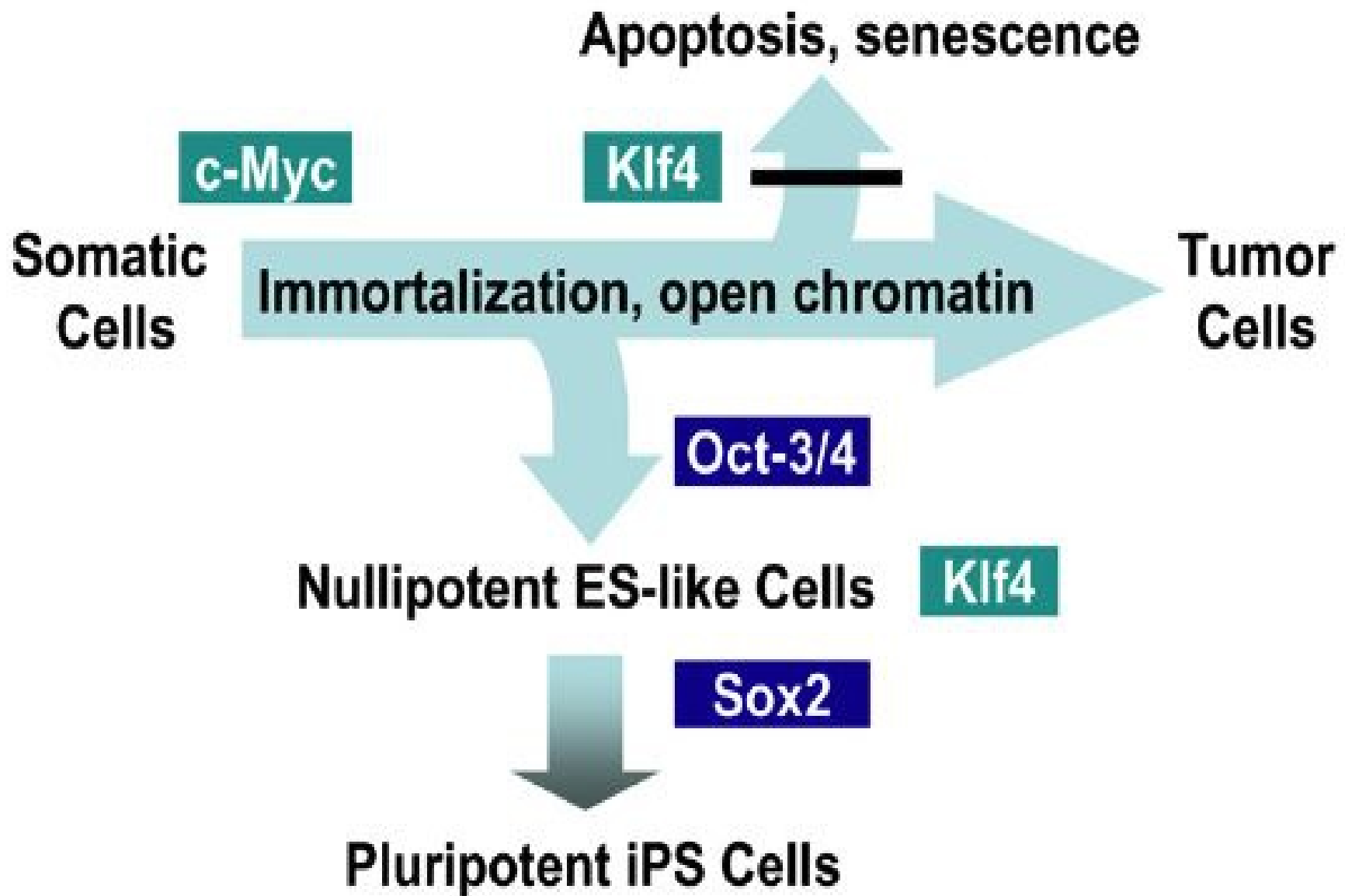


Five Factors Needed to Maintain Pluripotency

Table 1. Comparison of the Five Factors in the Phenotype of Loss-of-Function and Gain-of-Function Experiments

	Knockout ES Cells	Knockout Embryos	Overexpression in ES Cells
<i>Oct-3/4</i>	Cannot be established	No epiblast	Induces differentiation
	Niwa et al., 2000	Nichols et al., 1998	Niwa et al., 2000
<i>Sox2</i>	Cannot be established	No epiblast	Does not induce differentiation
	Masui et al., 2007	Avilion et al., 2003	Does not induce LIF independency
			M. Nakagawa and S.Y., unpublished data
<i>c-Myc</i>	Can be established	Normal epiblast	Does not induce differentiation
	Normal self-renewal		Induces LIF independency
	Davis et al., 1993	Davis et al., 1993	Cartwright et al., 2005
<i>KLF4</i>	Not reported	Normal epiblast	Does not induce differentiation
		Katz et al., 2002	Induces LIF independency
			Y. Tokuzawa, M. Nakagawa, and S.Y., unpublished data
<i>Nanog</i>	Can be established	No epiblast	Does not induce differentiation
	Spontaneous differentiation		Induces LIF independency
	Mitsui et al., 2003	Mitsui et al., 2003	Chambers et al., 2003; Mitsui et al., 2003

Induction of Pluripotent Stem Cells (iPS) from Somatic Stem Cells



Adipose Tissue Provides iPSC Efficiently

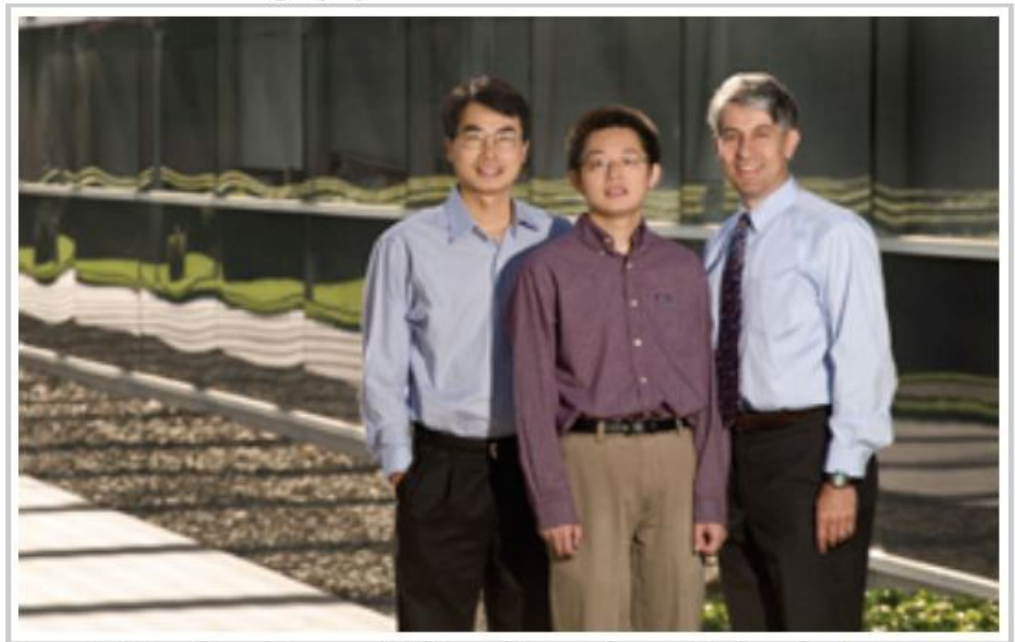
'Liposuction leftovers' easily converted to iPS cells, study shows

BY KRISTA CONGER

Globs of human fat removed during liposuction conceal versatile cells that are more quickly and easily coaxed to become induced pluripotent stem cells, or iPS cells, than are the skin cells most often used by researchers, according to a new study from Stanford's School of Medicine.

"We've identified a great natural resource," said Stanford surgery professor and co-author of the research, Michael Longaker, MD, who has called the readily available liposuction leftovers "liquid gold." Reprogramming adult cells to function like embryonic stem cells is one way researchers hope to create patient-specific cell lines to regenerate tissue or to study specific diseases in the laboratory.

Steve Fisch Photography



Joseph Wu, Ning Sun and Michael Longaker collaborated on research that showed stem cells found in fat tissue could easily be converted into iPS cells.

Using CRE – Recombinase to Remove Viral Transforming DNA from iPSCs

Parkinson's Disease Patient-Derived Induced Pluripotent Stem Cells Free of Viral Reprogramming Factors

Frank Soldner,^{1,4} Dirk Hockemeyer,^{1,4} Caroline Beard,¹ Qing Gao,¹ George W. Bell,¹ Elizabeth G. Cook,¹ Gunnar Hargus,³ Alexandra Blak,³ Oliver Cooper,³ Maisam Mitalipova,¹ Ole Isacson,³ and Rudolf Jaenisch^{1,2,*}

¹The Whitehead Institute, 9 Cambridge Center, Cambridge, MA 02142, USA

²Department of Biology, Massachusetts Institute of Technology, 31 Ames Street, Cambridge, MA 02139, USA

³Udall Parkinson Disease Research Center of Excellence, Center for Neuroregeneration Research, McLean Hospital/Harvard Medical School, Belmont, MA 02478, USA

⁴These authors contributed equally to this work

*Correspondence: jaenisch@wi.mit.edu

DOI 10.1016/j.cell.2009.02.013

Cre-Lox Recombination to Remove Viral DNA

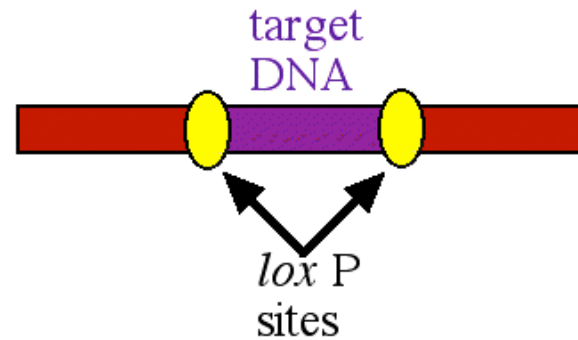


Figure 1. A pair of *lox P* sites (yellow ovals) flanking the target DNA (purple) to be deleted.

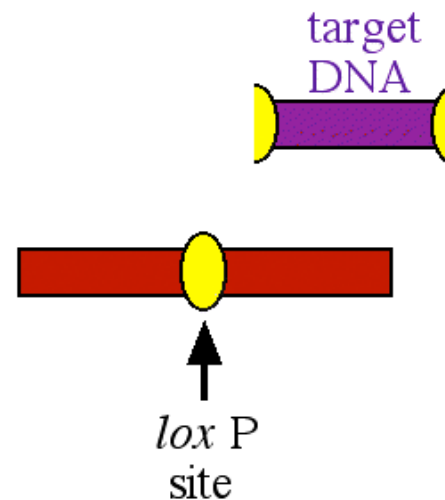


Figure 2. After the cre enzyme has excised the target DNA, one *lox P* site is left behind and the two flanking fragments of DNA are spliced together. The target DNA is excised and degraded.

Inducing iPSCs using Transcription Factor Proteins

Cell
PRESS

Cell Stem Cell
Brief Report

Generation of Human Induced Pluripotent Stem Cells by Direct Delivery of Reprogramming Proteins

Dohoon Kim,^{1,5} Chun-Hyung Kim,^{1,5} Jung-Il Moon,¹ Young-Gie Chung,³ Mi-Yoon Chang,¹ Baek-Soo Han,¹ Sanghyeok Ko,¹ Eungi Yang,¹ Kwang Yul Cha,⁴ Robert Lanza,^{3,*} and Kwang-Soo Kim^{1,2,4,*}

¹Molecular Neurobiology Laboratory, Department of Psychiatry and McLean Hospital, Harvard Medical School

²Harvard Stem Cell Institute

115 Mill Street, Belmont, MA 02478, USA

³Stem Cell and Regenerative Medicine International, 381 Plantation Street, Worcester, MA 01605, USA

⁴CHA Stem Cell Institute, CHA University, 606-16 Yoeksam 1-dong, Gangnam-gu, Korea

⁵These authors contributed equally to this work


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DOI 10.1016/j.stem.2009.05.005

Direct conversion of mouse fibroblasts to self-renewing, tripotent neural precursor cells

Ernesto Lujan^{a,b}, Soham Chanda^{a,c}, Henrik Ahlenius^{a,d}, Thomas C. Südhof^{c,e,1}, and Marius Wernig^{a,d,1}

^aInstitute for Stem Cell Biology and Regenerative Medicine, Departments of ^dPathology, ^bGenetics, and ^cMolecular and Cellular Physiology, and ^eHoward Hughes Medical Institute, Stanford University School of Medicine, Stanford, CA 94305



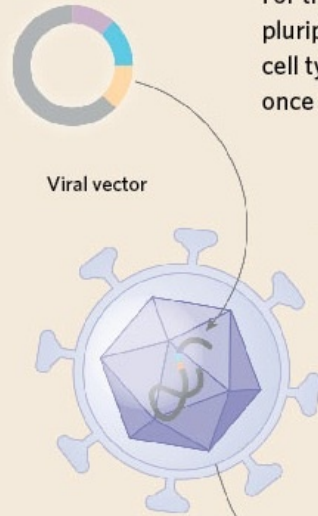
We recently showed that defined sets of transcription factors are sufficient to convert mouse and human fibroblasts directly into cells resembling functional neurons, referred to as “induced neuronal” (iN) cells. For some applications however, it would be desirable to convert fibroblasts into proliferative neural precursor cells (NPCs) instead of neurons. We hypothesized that NPC-like cells may be induced using the same principal approach used for generating iN cells. Toward this goal, we infected mouse embryonic fibroblasts derived from Sox2-EGFP mice with a set of 11 transcription factors highly expressed in NPCs. Twenty-four days after transgene induction, Sox2-EGFP⁺ colonies emerged that expressed NPC-specific genes and differentiated into neuronal and astrocytic cells. Using stepwise elimination, we found that Sox2 and FoxG1 are capable of generating clonal self-renewing, bipotent induced NPCs that gave rise to astrocytes and functional neurons. When we added the Pou and Homeobox domain-containing transcription factor Brn2 to Sox2 and FoxG1, we were able to induce tripotent NPCs that could be differentiated not only into neurons and astrocytes but also into oligodendrocytes. The transcription factors FoxG1 and Brn2 alone also were capable of inducing NPC-like cells; however, these cells generated less mature neurons, although they did produce astrocytes and even oligodendrocytes capable of integration into dysmyelinated Shiverer brain. Our data demonstrate that direct lineage reprogramming using target cell-type-specific transcription factors can be used to induce NPC-like cells that potentially could be used

Direct versus indirect Cell Reprogramming

<http://www.the-scientist.com/?articles.view/articleNo/39241/title/A-Twist-of-Fate/>

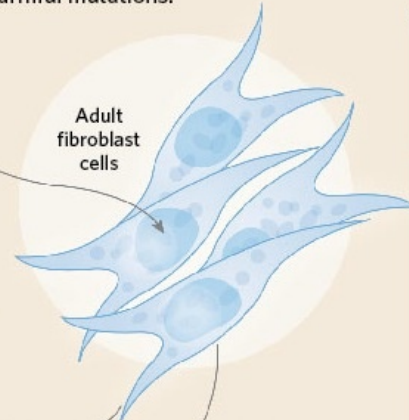
CELLULAR REPROGRAMMING

For the better part of the past decade, researchers have been reprogramming adult cell types, either into induced pluripotent stem cells (iPSCs), which themselves can give rise to diverse cell types, or directly into other differentiated cell types through a process called direct reprogramming. Such approaches support the switching of diverse cell types once believed to be permanently locked in their differentiated form.



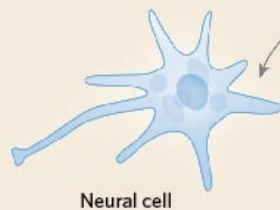
Viral vector

Traditionally, relevant transcription factors encoded by genetic material were carried by retro- or lentivirus vectors and integrated into the host cell genome. More recently, the use of nonintegrating vectors, RNA, or small molecules have been developed to minimize the chance of harmful mutations.

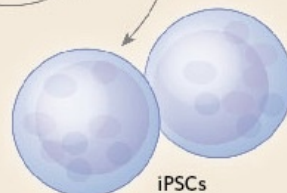


Adult fibroblast cells

Fibroblasts were the first and remain the most common type of cell to be reprogrammed, but other cells, such as lymphocytes, which can be isolated from blood, are also proving to be successful starting points for stem-cell generation.



Neural cell



iPSCs

Direct reprogramming into another adult cell type

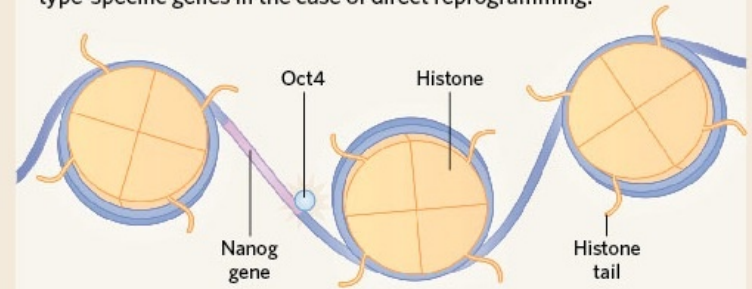
OR

Dedifferentiation into a pluripotent state

© LUCY READING-IKKANDA

OPEN CHROMATIN

Transfected transcription factors, such as Oct4, induce the expression of pluripotency-related genes, such as *Nanog*, or cell-type-specific genes in the case of direct reprogramming.

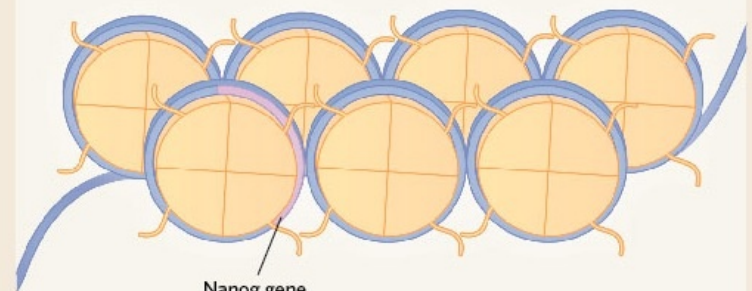


Nanog gene

Histone tail

CLOSED CHROMATIN

Sequences from pioneer factors, such as the myogenic factor MyoD, are also employed to increase reprogramming efficiency in the face of closed chromatin, which can inhibit access of the transfected transcription factors to their target genes.

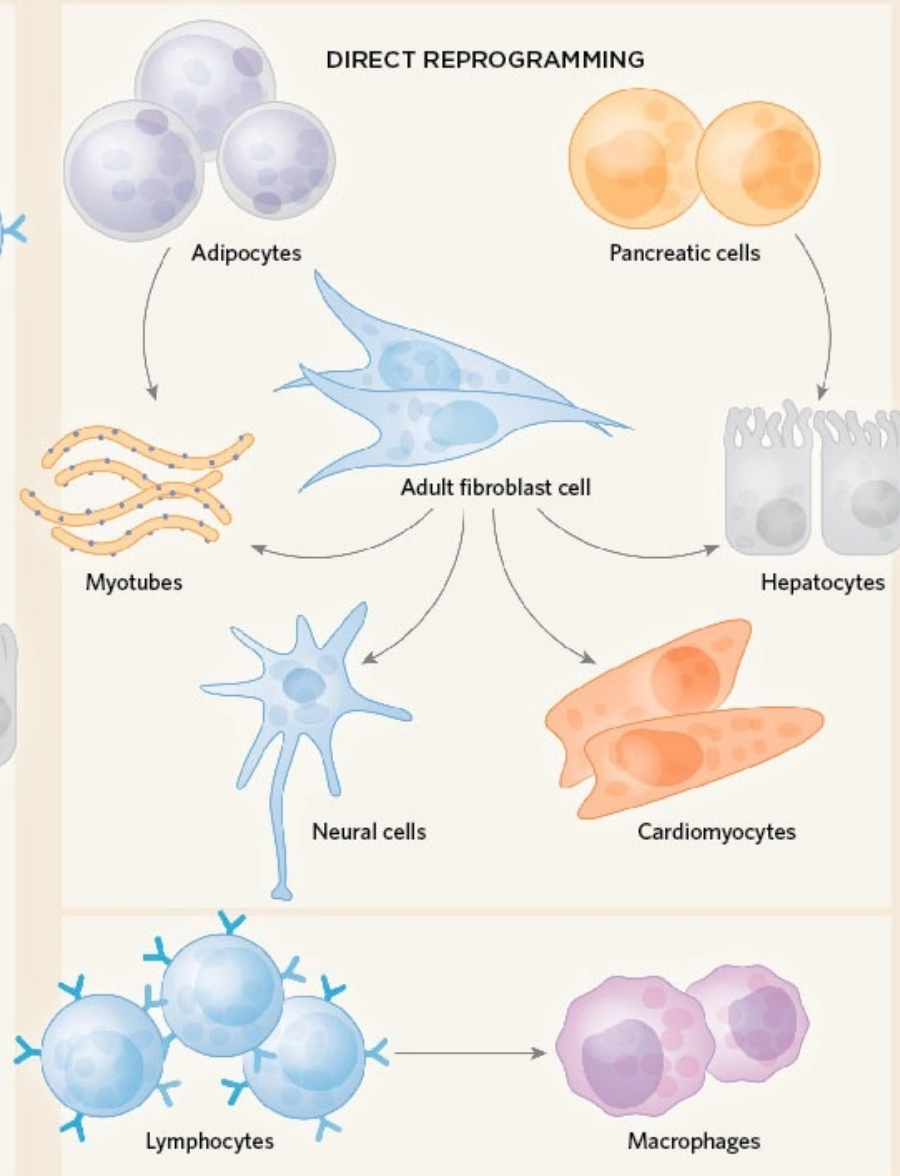
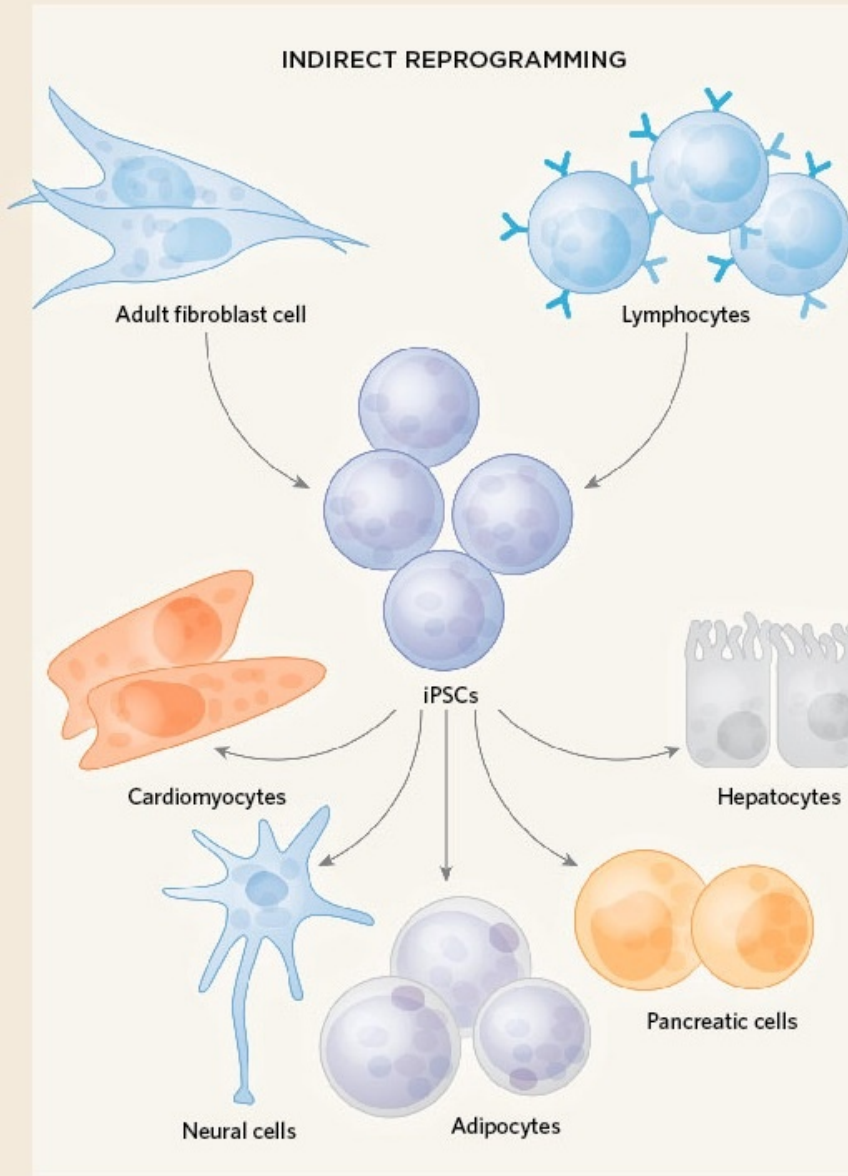


Nanog gene

<http://www.the-scientist.com/?articles.view/articleNo/39241/title/A-Twist-of-Fate/>

Cell Reprogramming *in vivo* & *in vitro*

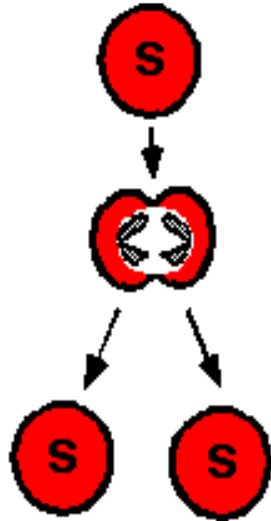
<http://www.the-scientist.com/?articles.view/articleNo/39241/title/A-Twist-of-Fate/>



<http://www.the-scientist.com/?articles.view/articleNo/39241/title/A-Twist-of-Fate/>

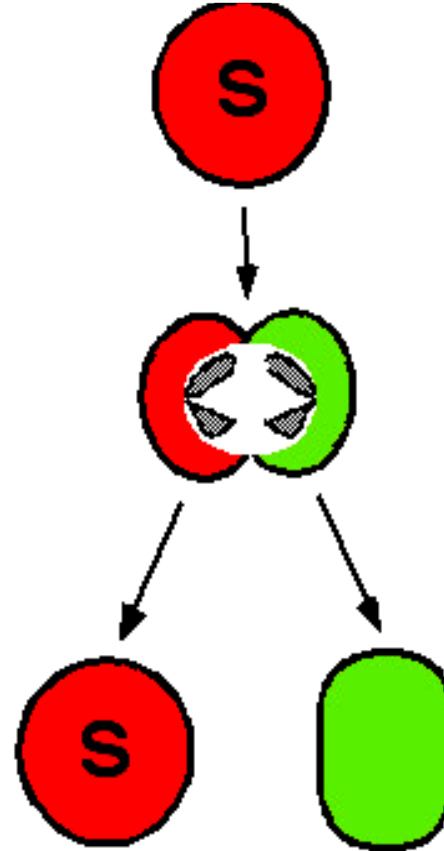
Alternate Stem Cell Fates

Embryonic
Stem Cells



stem cell
proliferation

Adult
Stem Cells



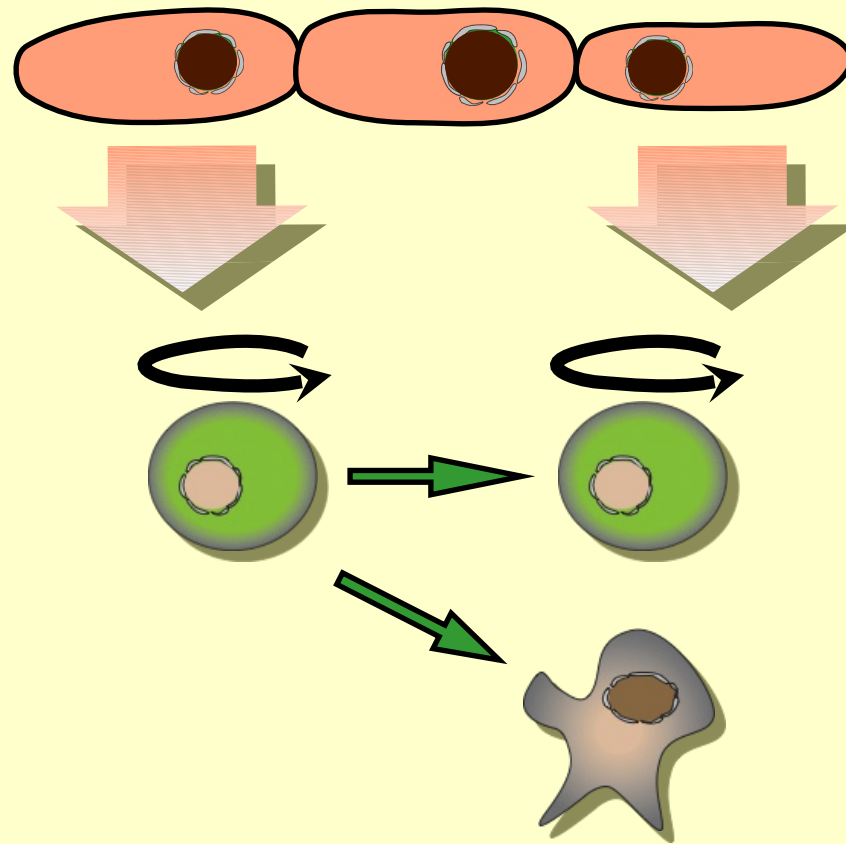
asymmetric
division

Adult
Stem Cells

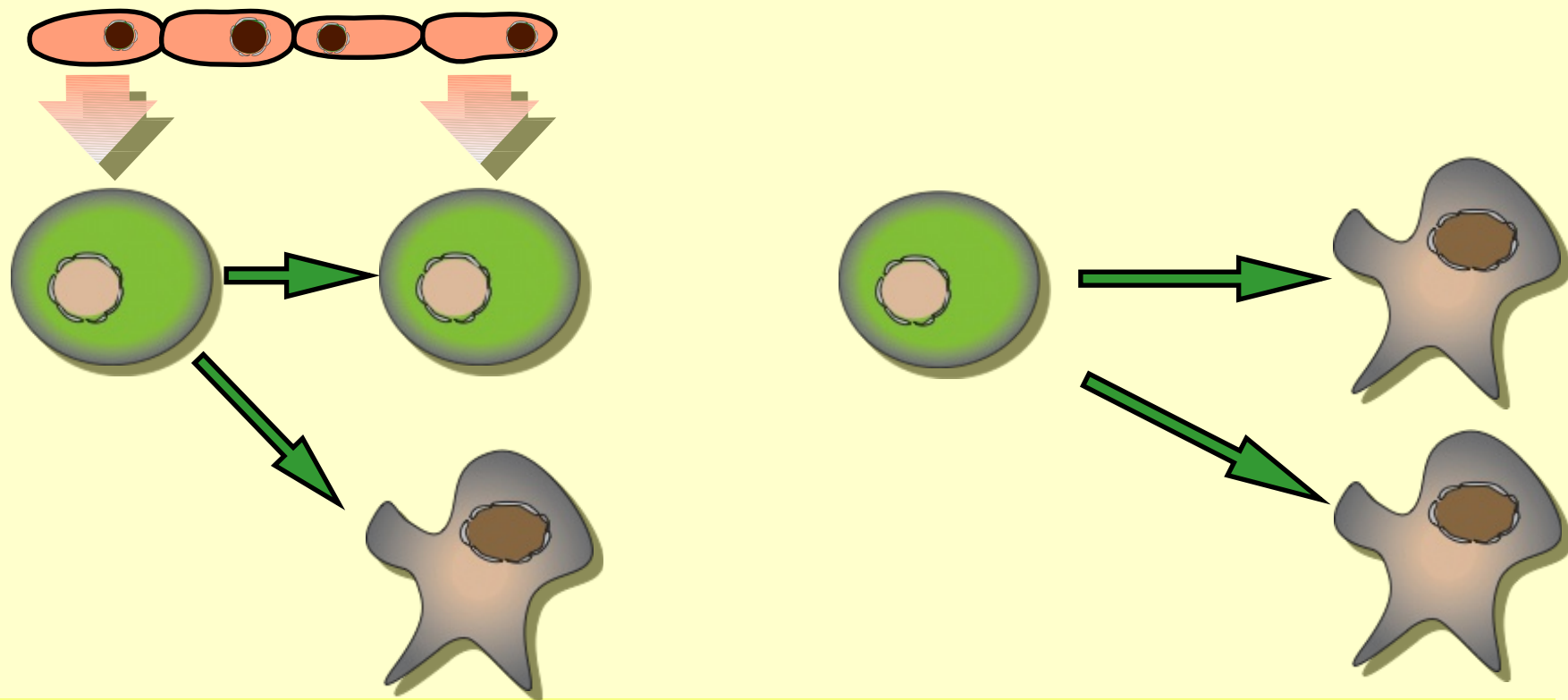


stem cell
loss

signals from niches maintain adult stem cells and tissues

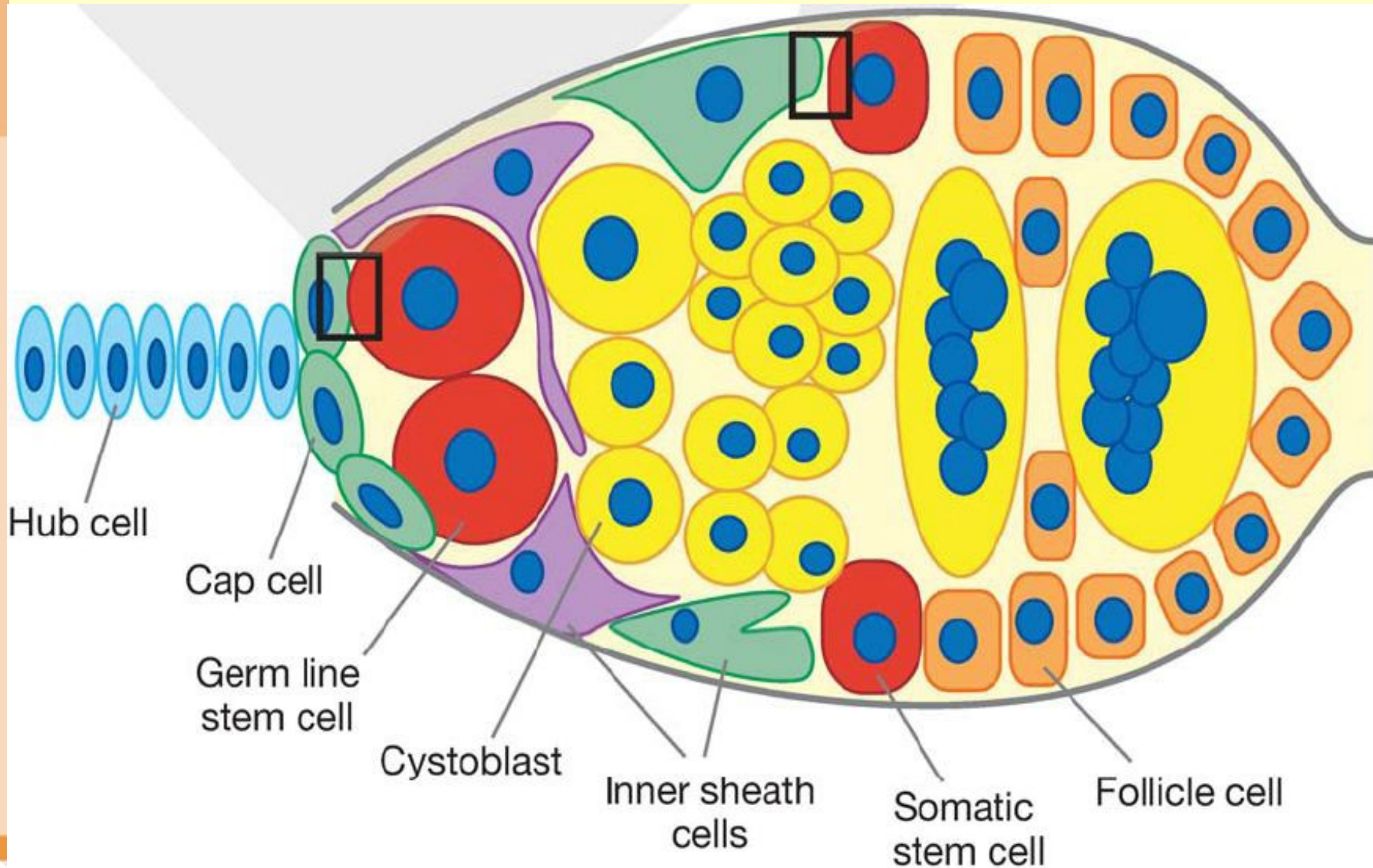


In the absence of niche signals, adult stem cells will differentiate, by default

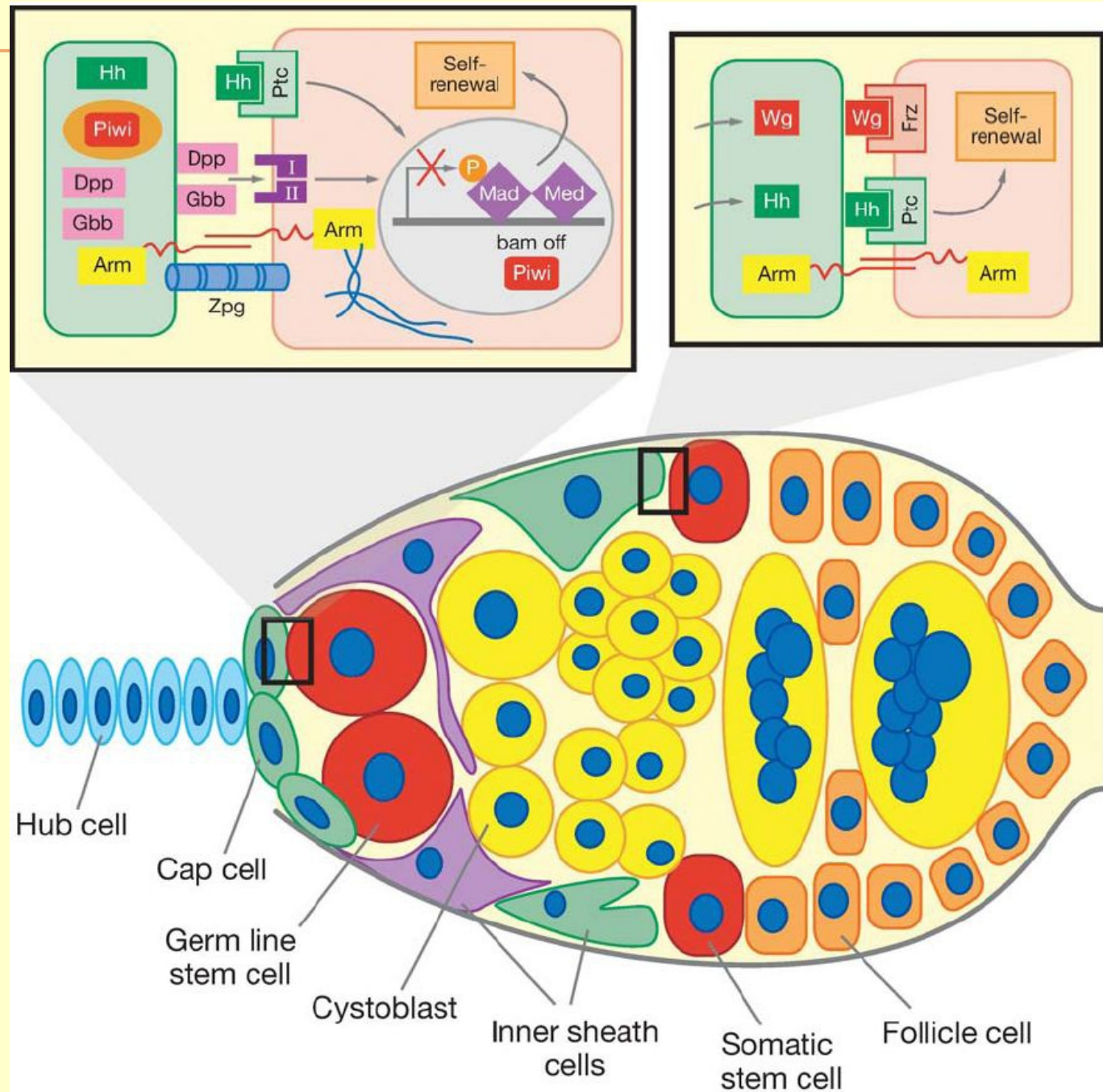


1. Self-renewal is proliferation coupled to blocking differentiation, controlled by signals.
2. Signals are local; niches have a limited capacity and cells compete for the signals
3. The signals control tissue homeostasis, also after damage

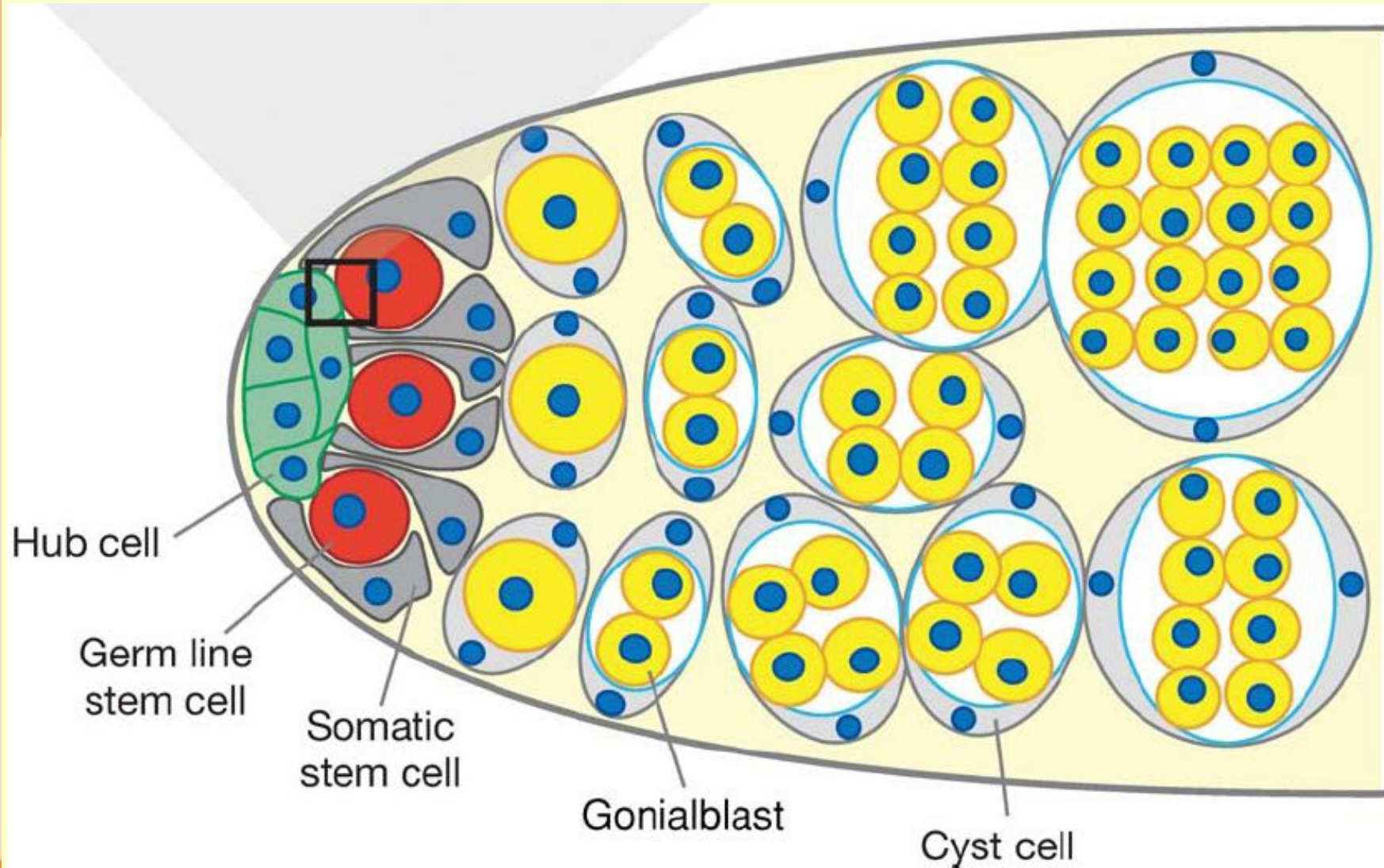
Oocyte Niche in the Drosophila Germarium



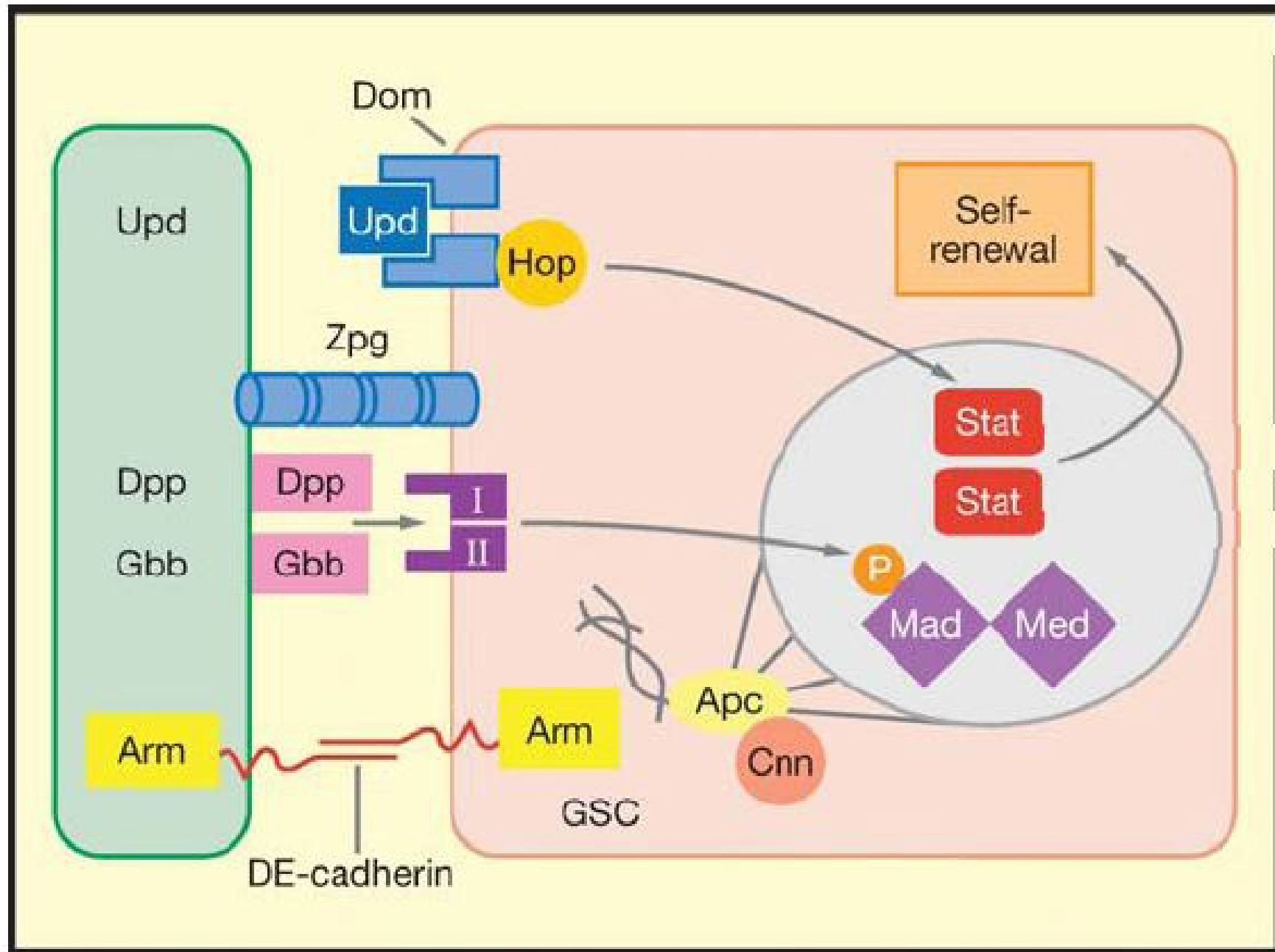
Cell-Cell Interactions at Oocyte Niche



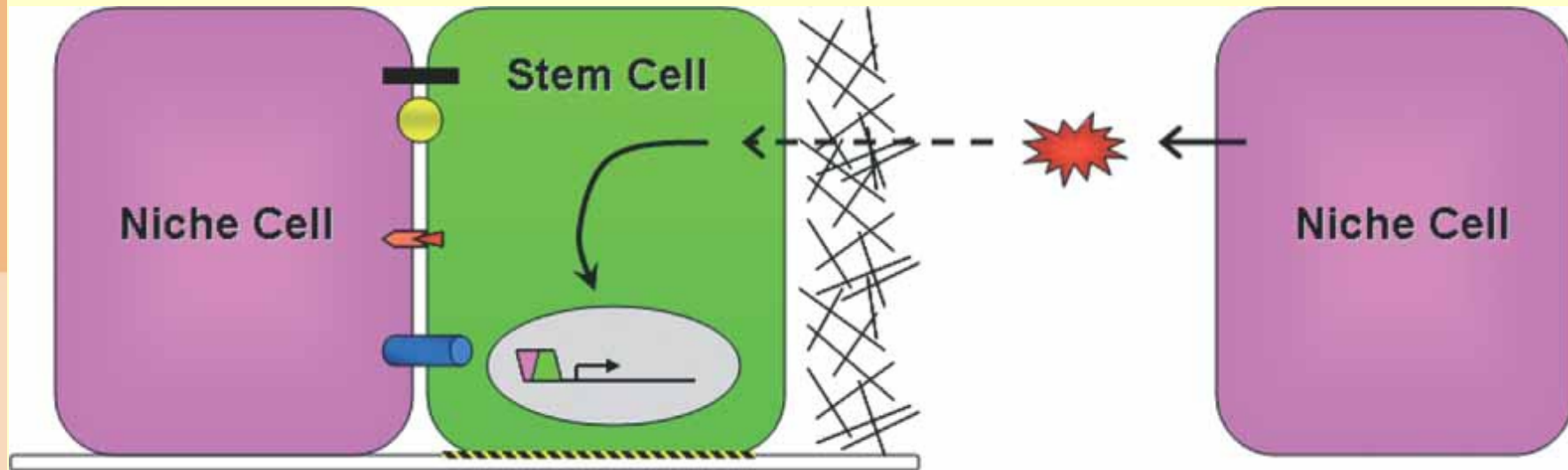
Drosophila Spermatogonial Niche









Cell-Cell Interactions at the Spermatogonial Niche






Summary of Stem Cell Niche Signals



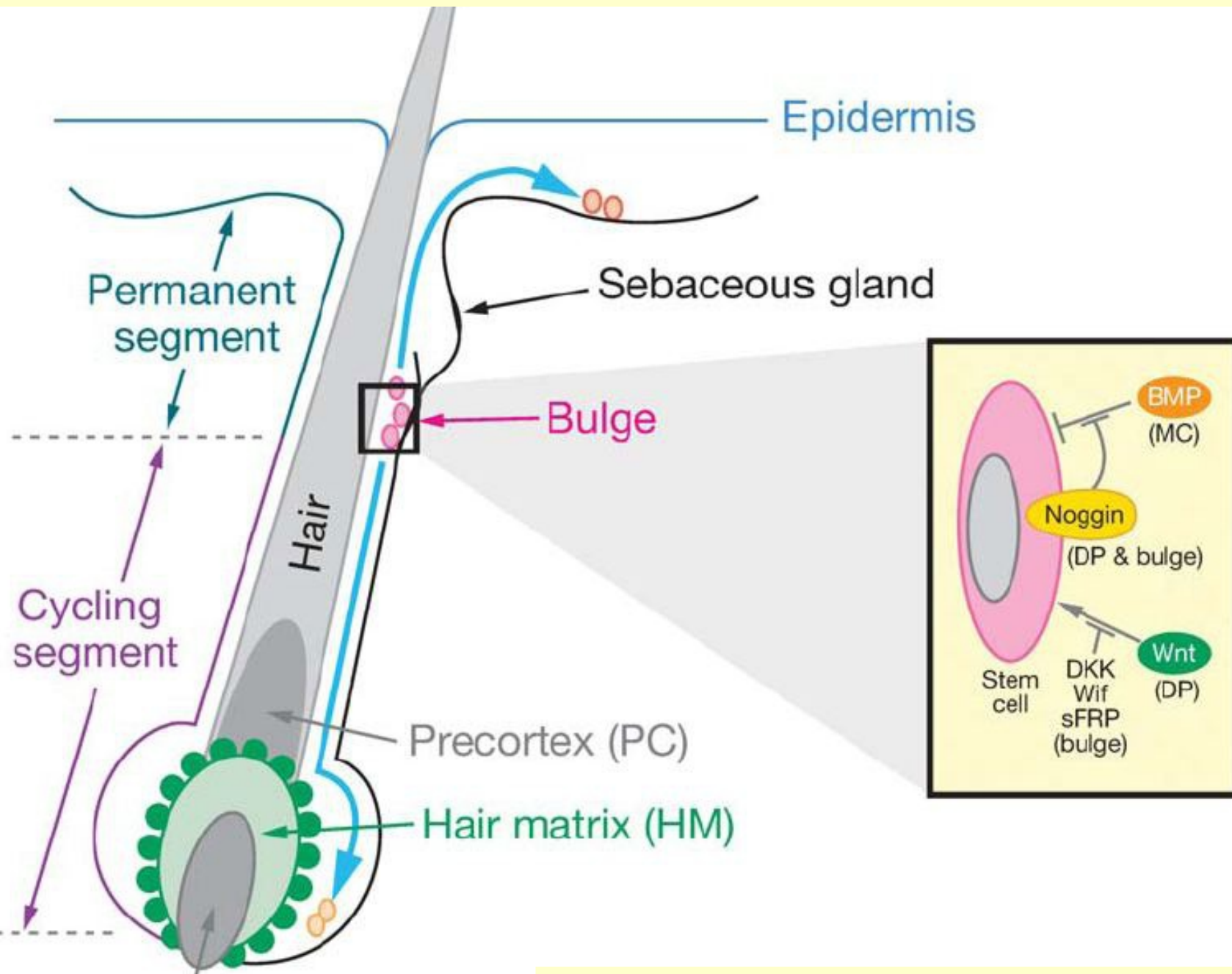
Physical Contact

-  **Tight Junction**
N, I
-  **Adherens Junction**
D, N
-  **Notch Signaling**
C, N, H, I
-  **Gap Junction**
D
-  **Basement Membrane**
N, E, I
-  **Extracellular Matrix**
D, N,

Diffusible Factors

-  **Pathway**
Wnt: C, E, H, I
BMP: D, N, E, I
JAK/STAT: D
Growth Factors: N
Hedgehog: I
PGE2: I
O₂: H
-  **Transcription Factor Activation**
-  **Signal Transduction**

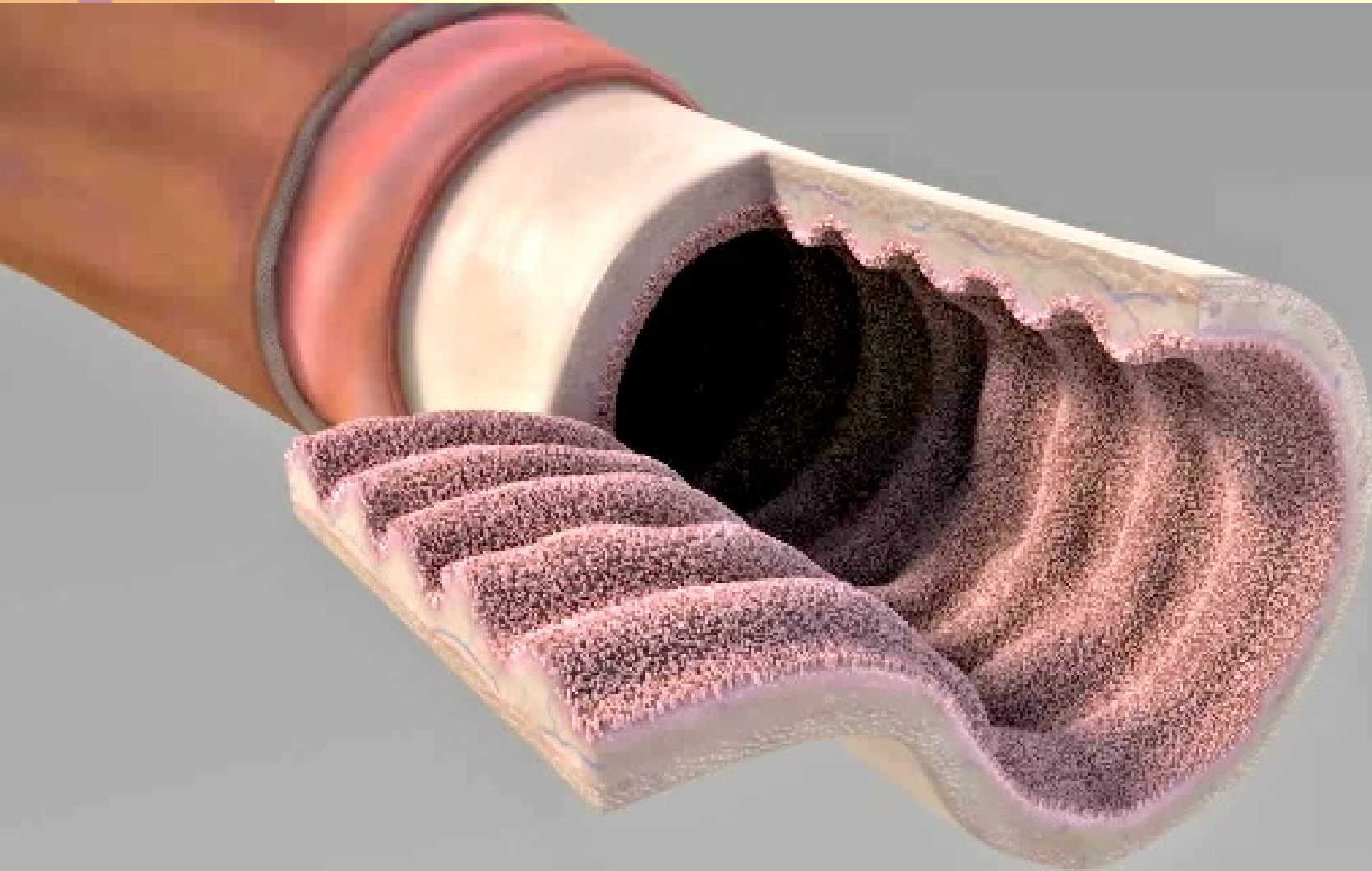
Hair Follicle Niche



Intestinal Stem Cells in Crypts

Clevers Lab|Digizyme

Rainbow Villi

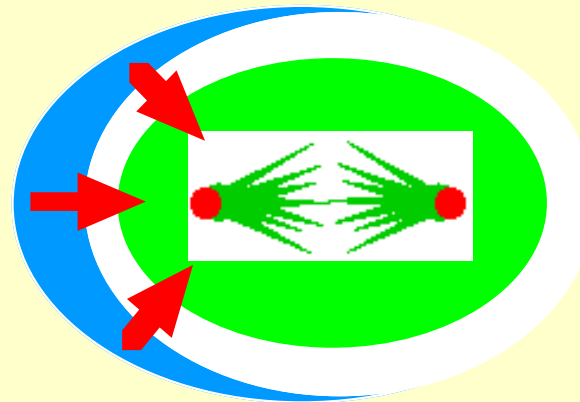


Clevers Lab | ANATOMY 3D

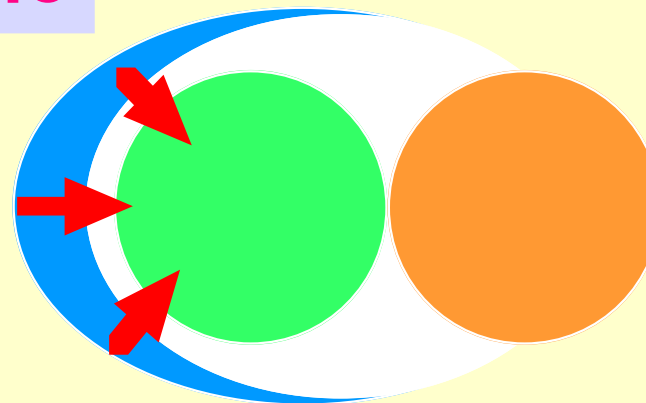


Asymmetric stem cell divisions

Extrinsic
factor(s)



Niche



John Cairns: The Immortal Parental Strands

Nature Vol. 255 May 15 1975

197

review article

Mutation selection and the natural history of cancer

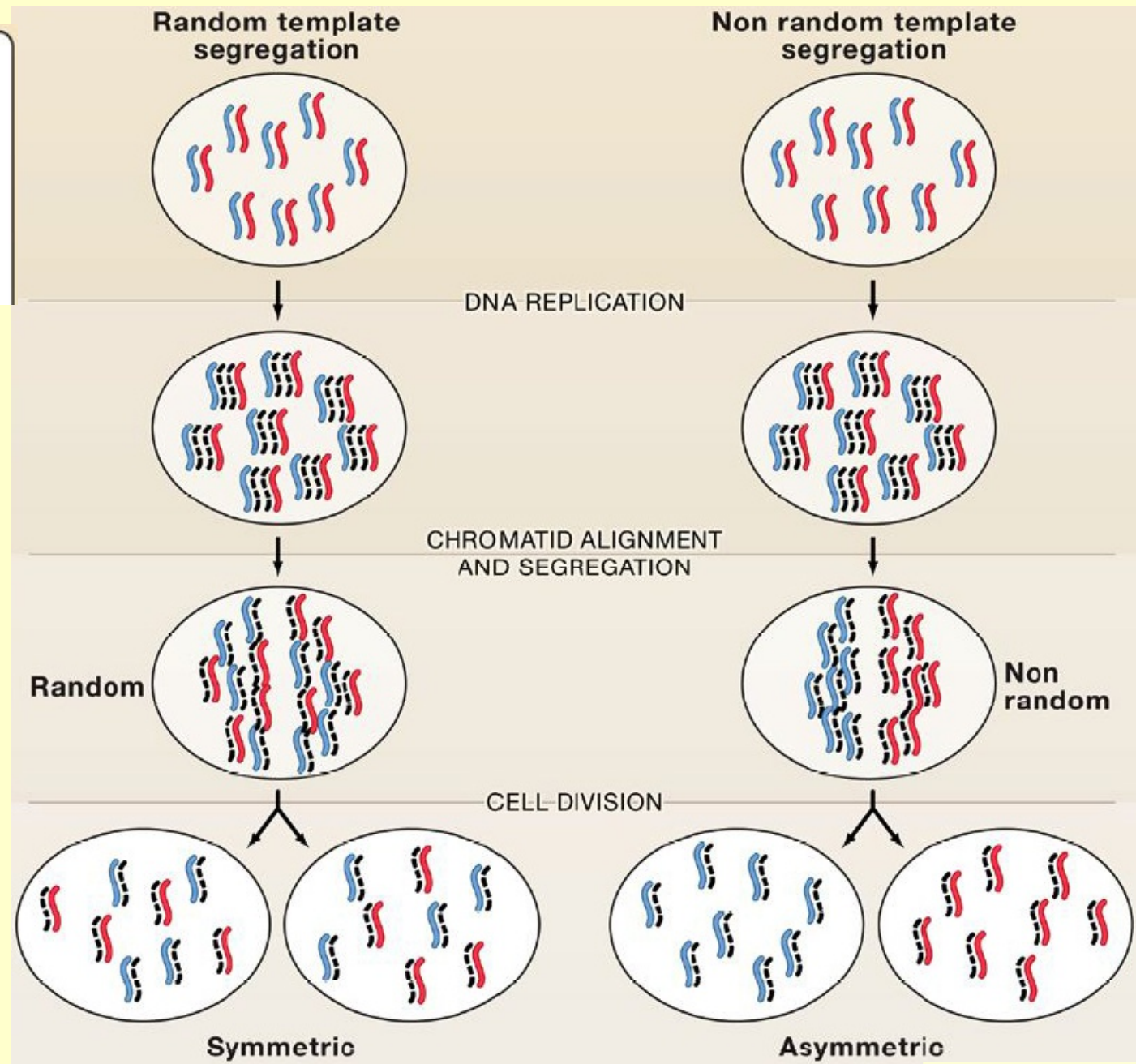
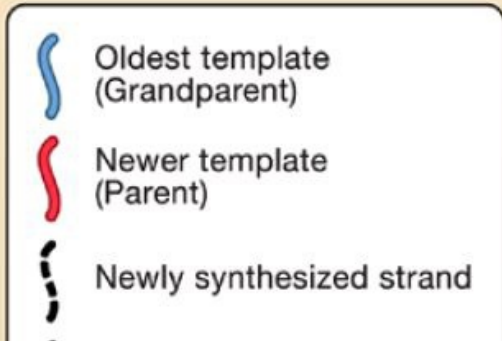
John Cairns*

Survival of the rapidly renewing tissues of long-lived animals like man requires that they be protected against the natural selection of fitter variant cells (that is, the spontaneous appearance of cancer). This article discusses three possible protective mechanisms and shows how they could explain various features of the natural history of certain common cancers of man.

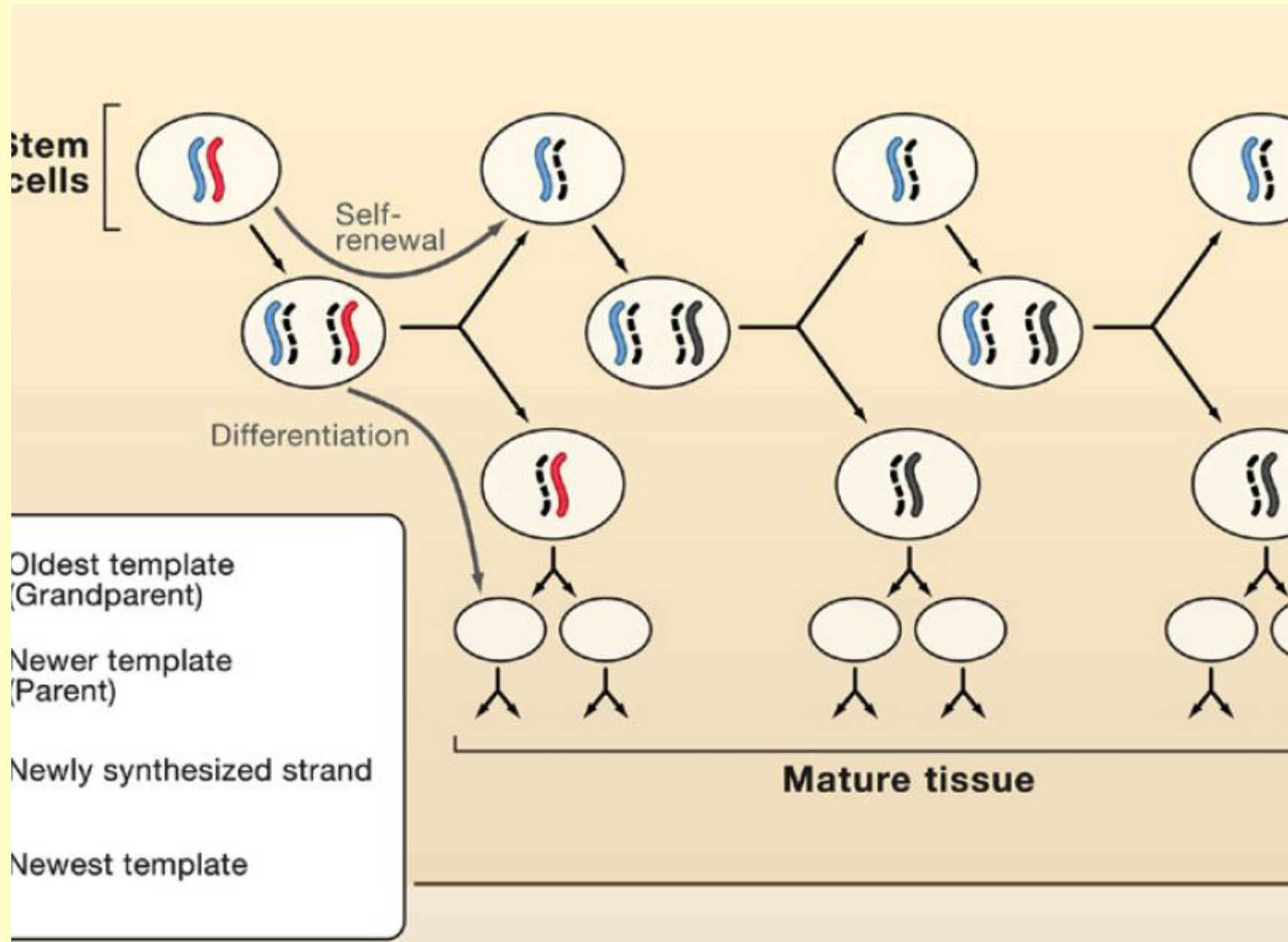
Motivation for Asymmetric Strand Segregation

- Adult rat contains 6×10^{10} cells
- In its small intestine, a rat sheds over 10^{13} epithelial cells during its lifetime.
- Requires 10^3 symmetric cell doublings from embryo to adult followed by 10^{13} asymmetric cell doublings during its lifetime
- How do epithelial cells minimize mutations that lead to cancer?

Asymmetric Segregation of Parental DNA Strands

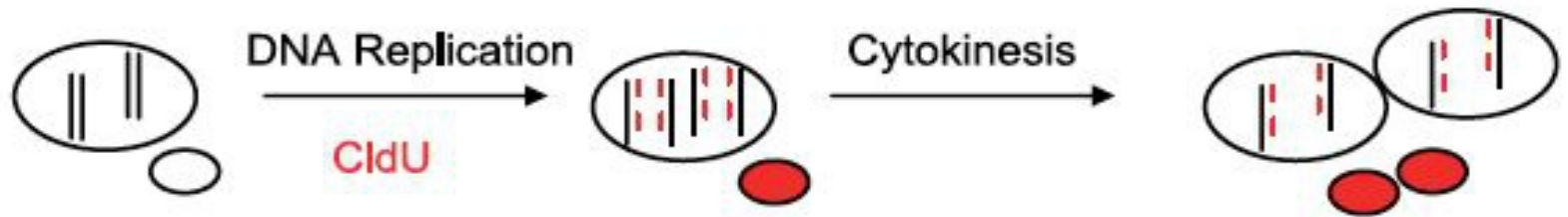


Asymmetric Stem Cell Growth with Asymmetric Parental Strand Segregation

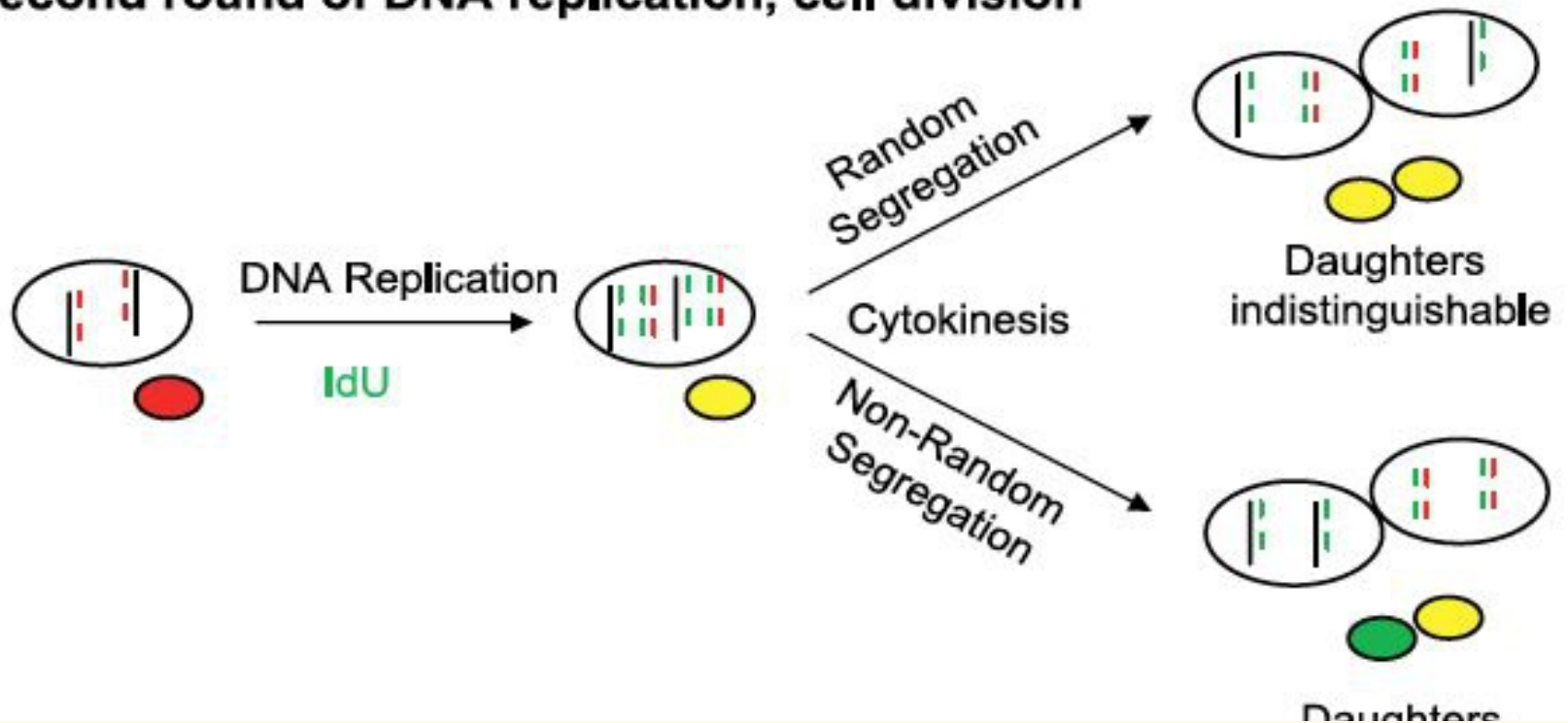


Asymmetric DNA Labeling Patterns

First round of DNA replication, cell division



Second round of DNA replication, cell division



Duplicating Muscle Cell Pairs Display Asymmetric DNA Labeling Patterns

B

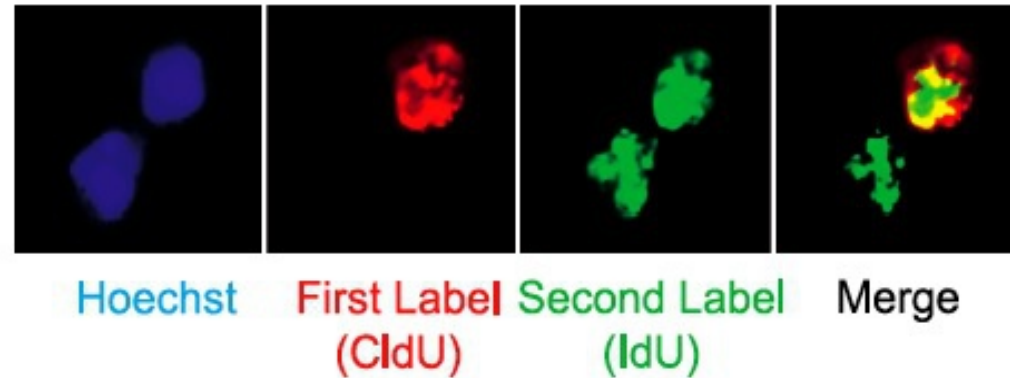
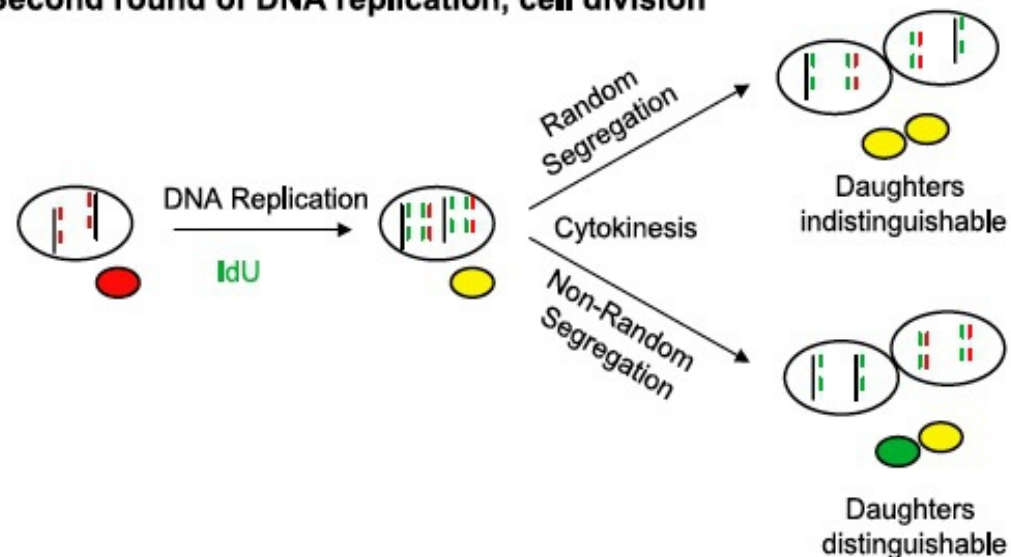


Figure 2. Evidence of Co-Segregation of DNA Template Strands during Muscle Progenitor Cell Division

(B) Cell pairs were immunostained for CldU and IdU. Shown is a representative photograph of an immunostained pair of cells, in which both daughter cells were labeled with the second label, IdU (green), but only one daughter inherited the first label, CldU (red).

Second round of DNA replication, cell division



Asymmetric Stem Cell Growth with Asymmetric Parental Strand Segregation

